

# The effects of storage on blood products

**Steven L. Spitalnik, M.D.**

**Laboratory of Transfusion Biology**



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*College of Physicians  
and Surgeons*

 **NewYork-Presbyterian**  
The University Hospitals of Columbia and Cornell

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# **The influence of donor genetics, diet, and environment on red blood cell storage quality**

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# Potential Conflicts of Interest

**Hemanext**

**Advisory Board**

**Tioma, Inc**

**Consultant**

**Team Conveyer  
Intellectual Prop.**

**Consultant**

**Ferrous Wheel Consultants**

**CEO**

**Worldwide Initiative  
For Rh Disease  
Eradication (WIRhE)**

**Executive  
Director**

# Laboratory of Transfusion Biology

## Columbia University



**I. Akpan**



**G. Brittenham**



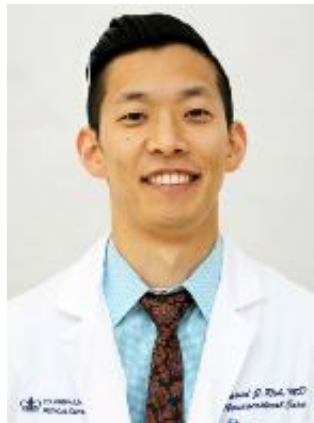
**R. Francis**



**E. Hod**



**K. Hudson**



**D. Roh**



**S. Spitalnik**



**T. Thomas**



**S. Weisberg**

# **Central Thesis**

**Blood products → Pharmaceuticals**

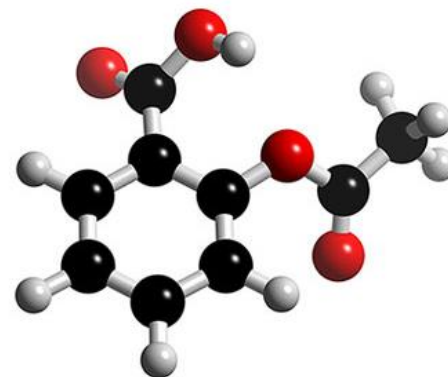
# **Central Thesis**

**Blood products → Pharmaceuticals**

**Complex biological products → “Pure drugs”**



# Aspirin = Acetylsalicylic acid



# **Medical Model of a Pharmaceutical**

**Active ingredient(s)**

**Purity**

**Inactive ingredients: binders, fillers, etc.**

**Stability: shelf life, expiration date, storage conditions**

**Dosages**

**Route of administration, Bioavailability**

**Pharmacokinetics/Pharmacodynamics**

**Indications**

**Clinical effectiveness**

**Adverse outcomes**

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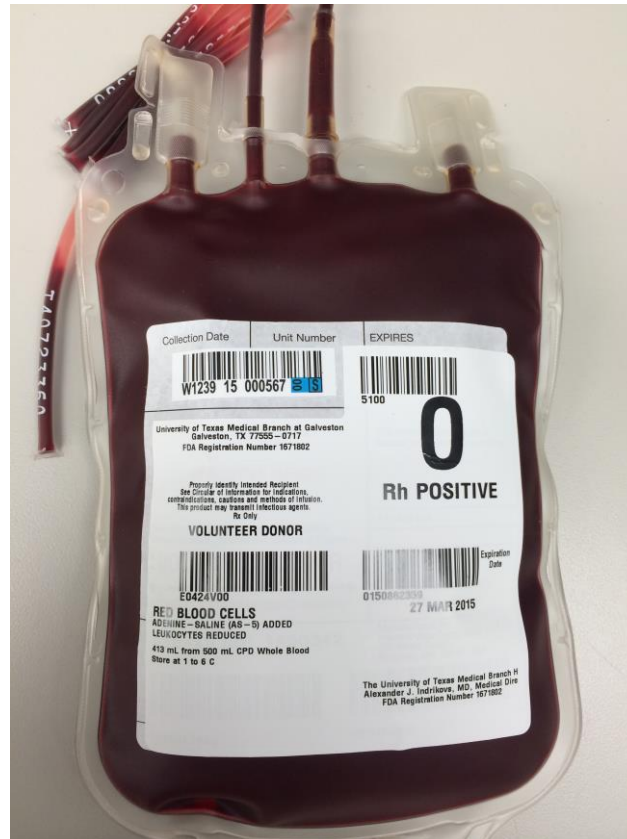
**Clinical effectiveness**

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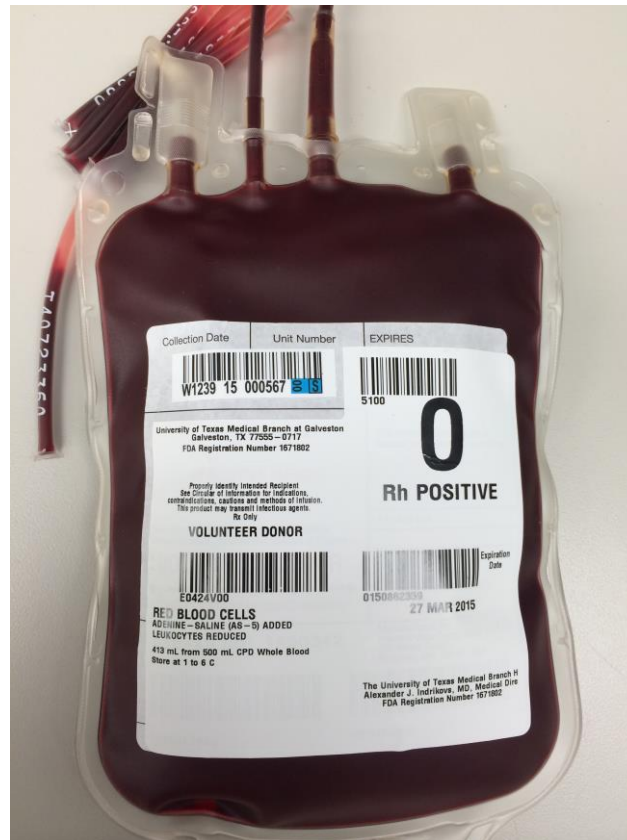
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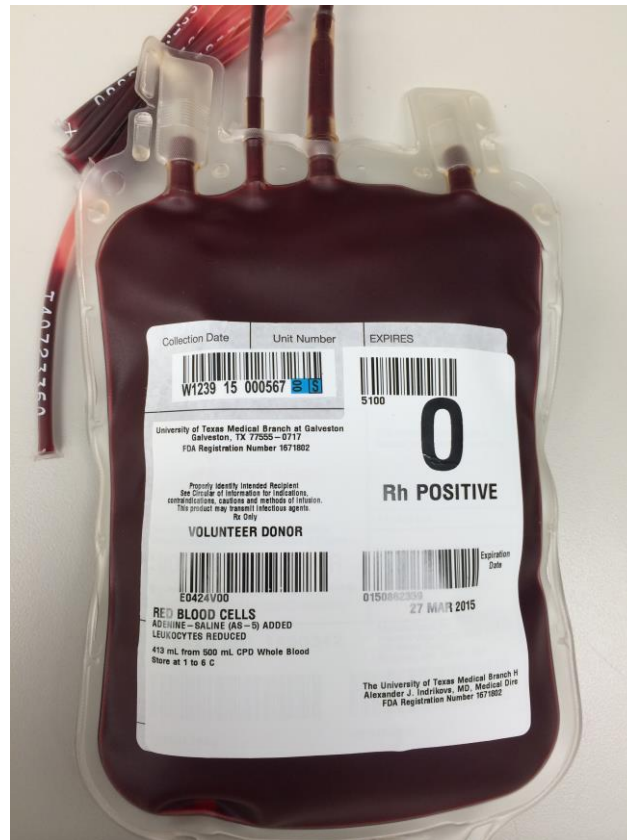


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## Blood “Pharm-ing”?

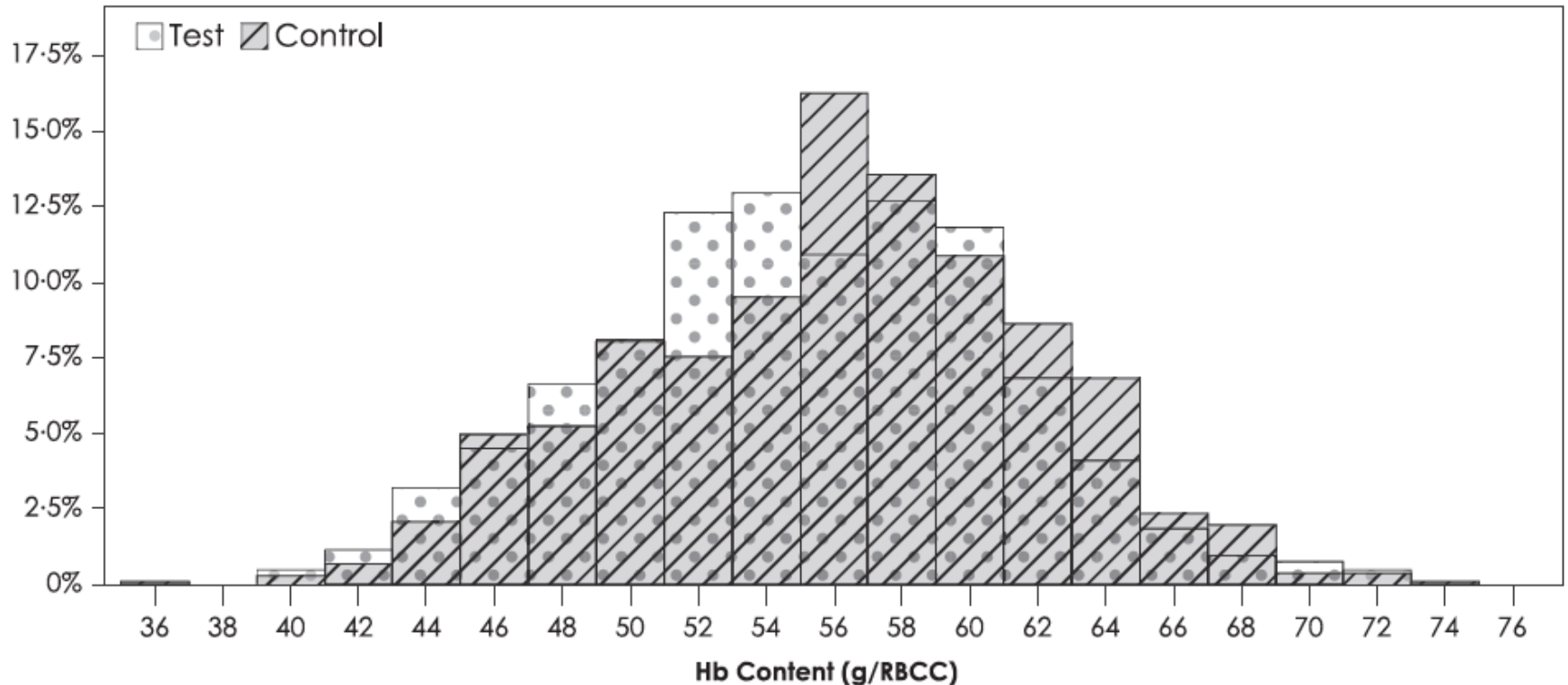
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## What about better starting material?



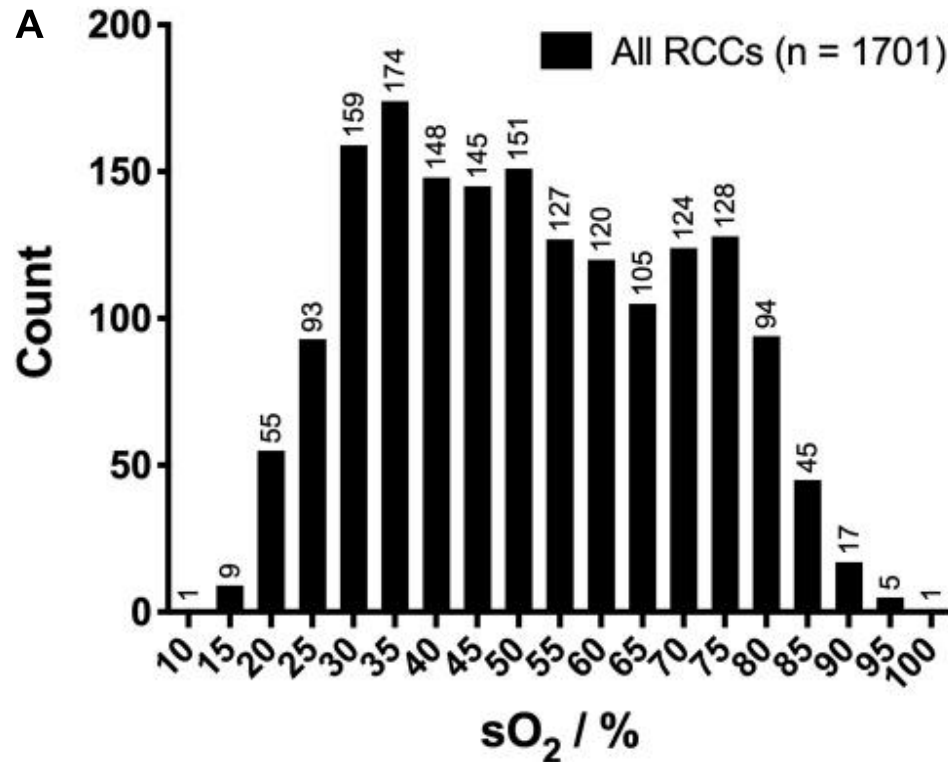
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Aydinok et al. Brit. J. Haematol. 186:625-636, 2019

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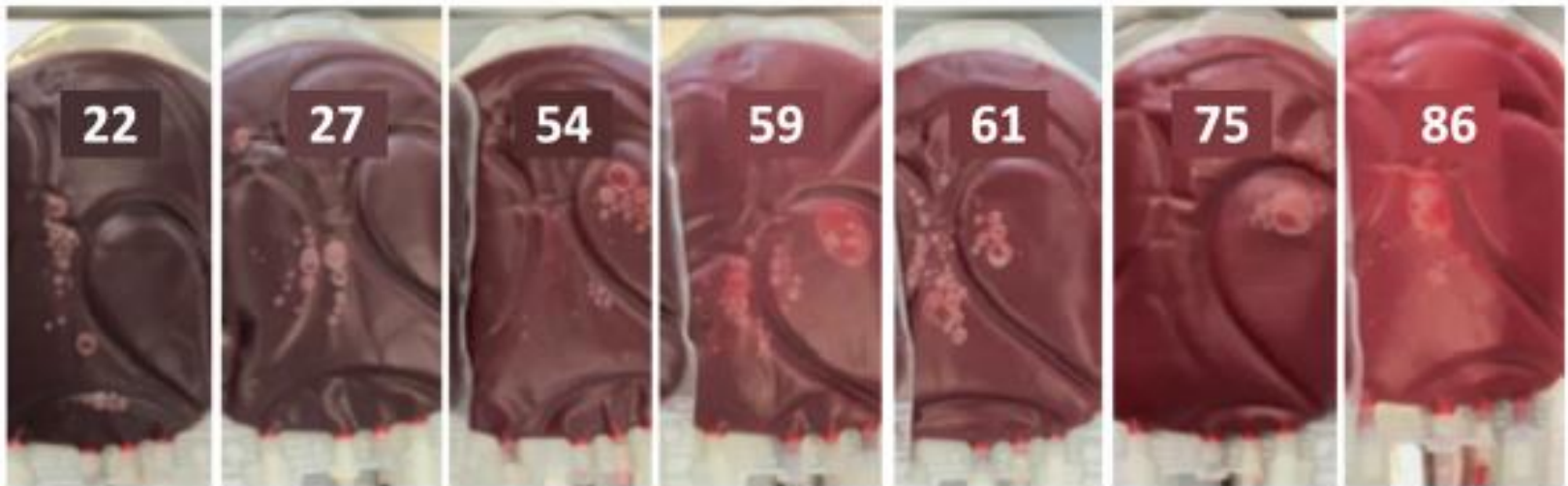
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Bardyn et al. Front. Physiol. 11:616457, 2020

## What about better starting material?

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**Bardyn et al. Front. Physiol. 11:616457, 2020**

**What about better starting material?**

**What do we know about  
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**Infuse free hemoglobin, etc.**

# What do we know about refrigerator-stored RBCs?

As storage time increases (FDA criteria):

Increasing hemolysis *ex vivo* (<1.0%)

**Infuse free hemoglobin, etc.**

Decreasing 24-hr post-transfusion recovery *in vivo* ( $\sim \geq 75\%$ )

**Less than optimal dose**

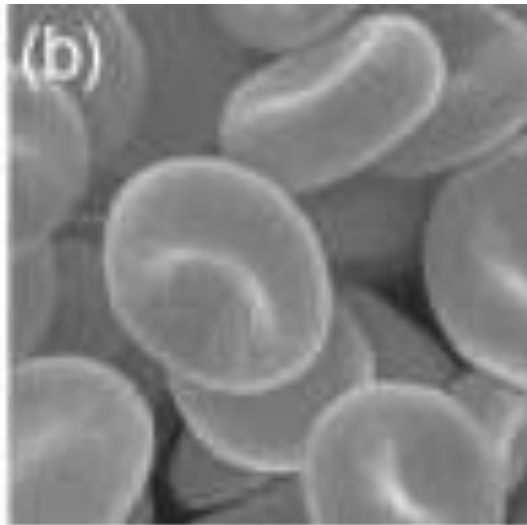
**What happens to the RBCs  
during refrigerated storage?**



# The “RBC storage lesion”

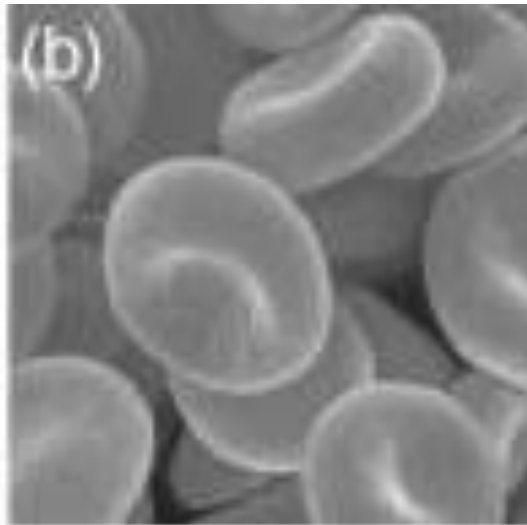
- ↓ 2,3-DPG, GSH, ATP
- ↓ Nitric oxide
- ↑ Protein oxidation
- ↑ Membrane- & cytoskeletal-associated hemoglobin
- ↑ Membrane lipid peroxidation
- ↑ Lysophosphatidylcholine species
- ↑ Vesiculation and membrane loss
- ↓ Deformability
- ↑ Phosphatidylserine exposure
- ↓ CD47

# The “RBC storage lesion”

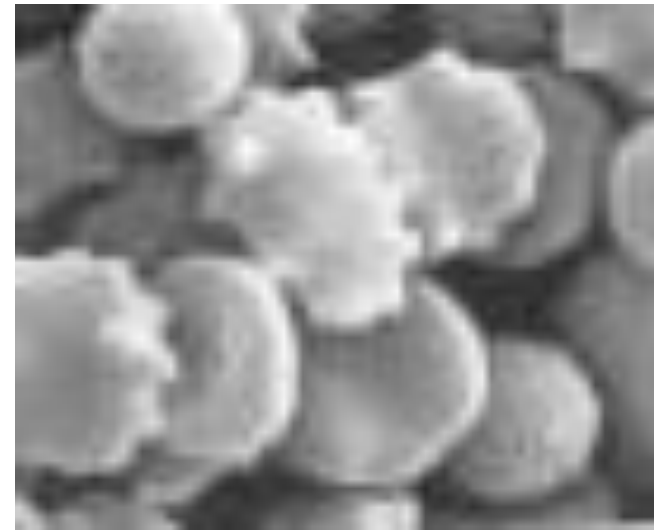


**Day 0**

# The “RBC storage lesion”



**Day 0**



**Day 42**

# The “RBC storage lesion”

↓ 2,3-DPG, GSH, ATP

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↓ CD47

# The “RBC storage lesion”

## Final common pathway?

Metabolic dysfunction & oxidative stress →

↓ Deformability

↑ “Eat me” signals (phosphatidylserine)

↓ “Don’t eat me” signals (CD47)

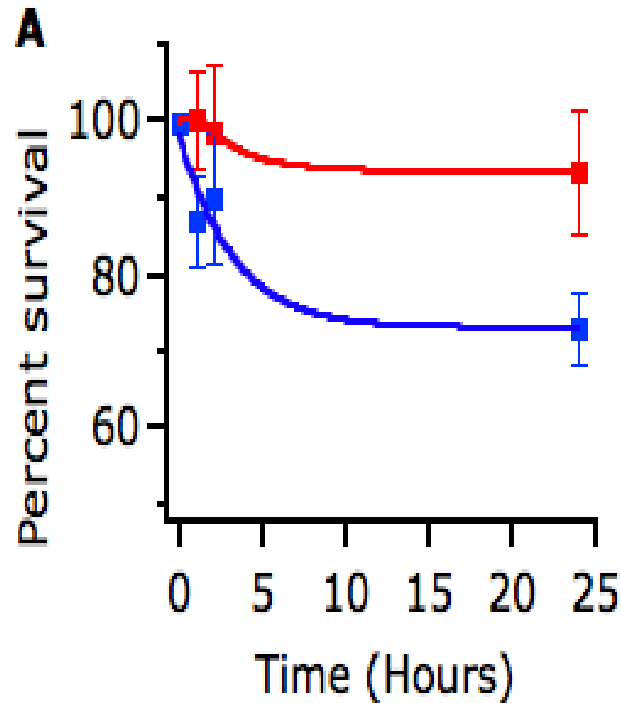
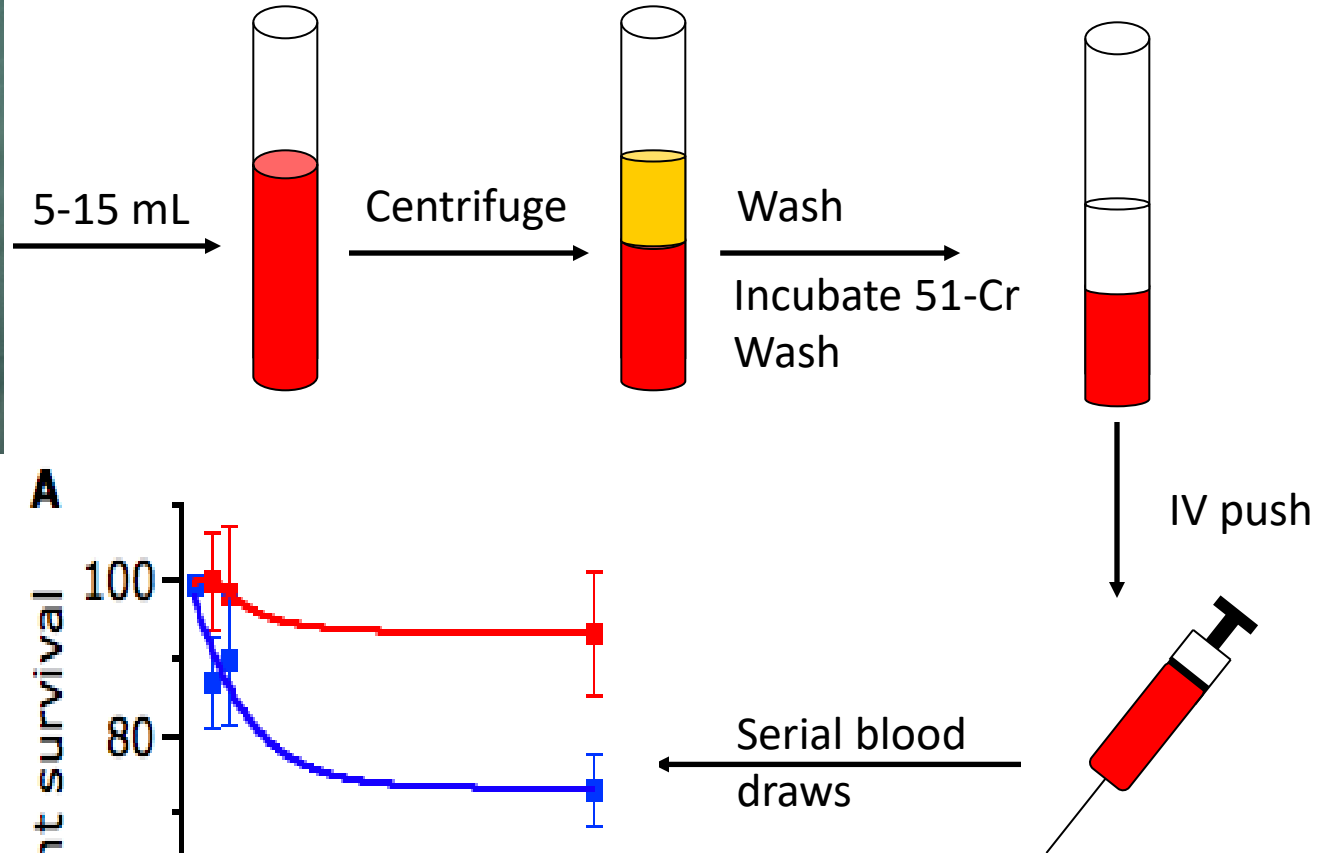
↑ Hemolysis *in vitro*

↑ RBC clearance *in vivo*

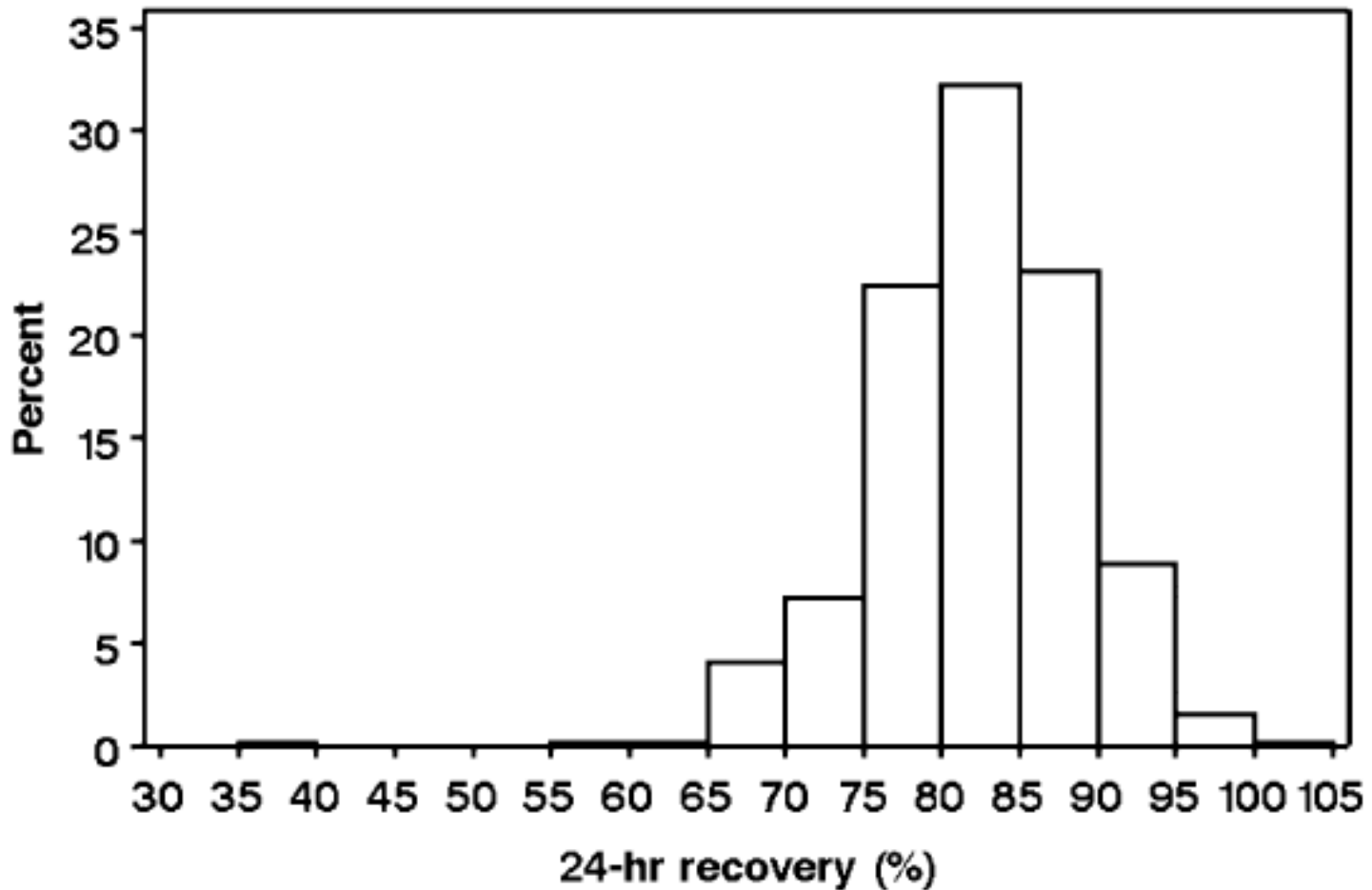
Intravascular and extravascular hemolysis

# **RBC Clearance Variability**

# RBC Recovery Study



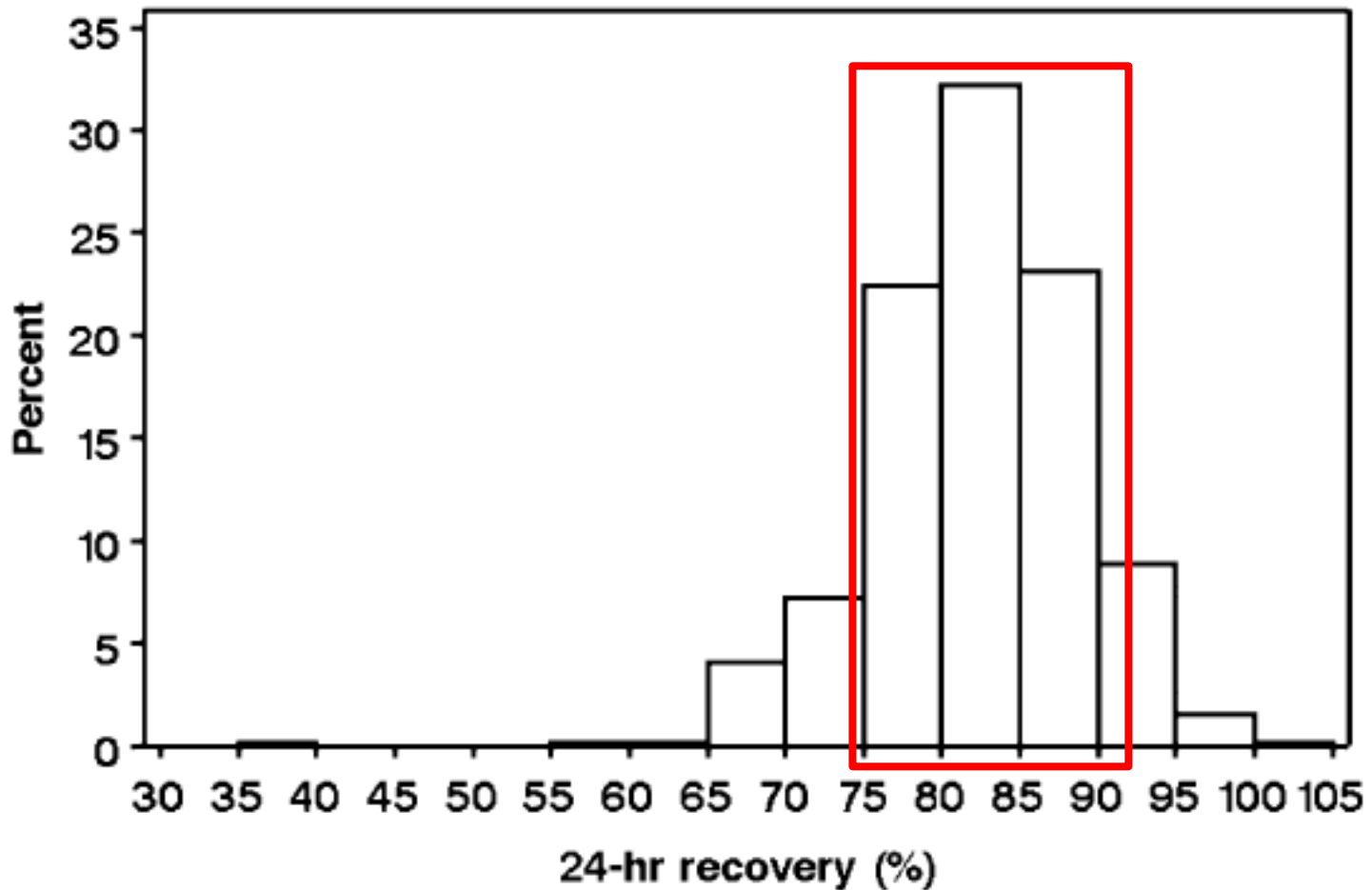
# 24-hr RBC recovery in 641 healthy volunteers



Dumont et al. Transfusion 48:1053-60, 2008.

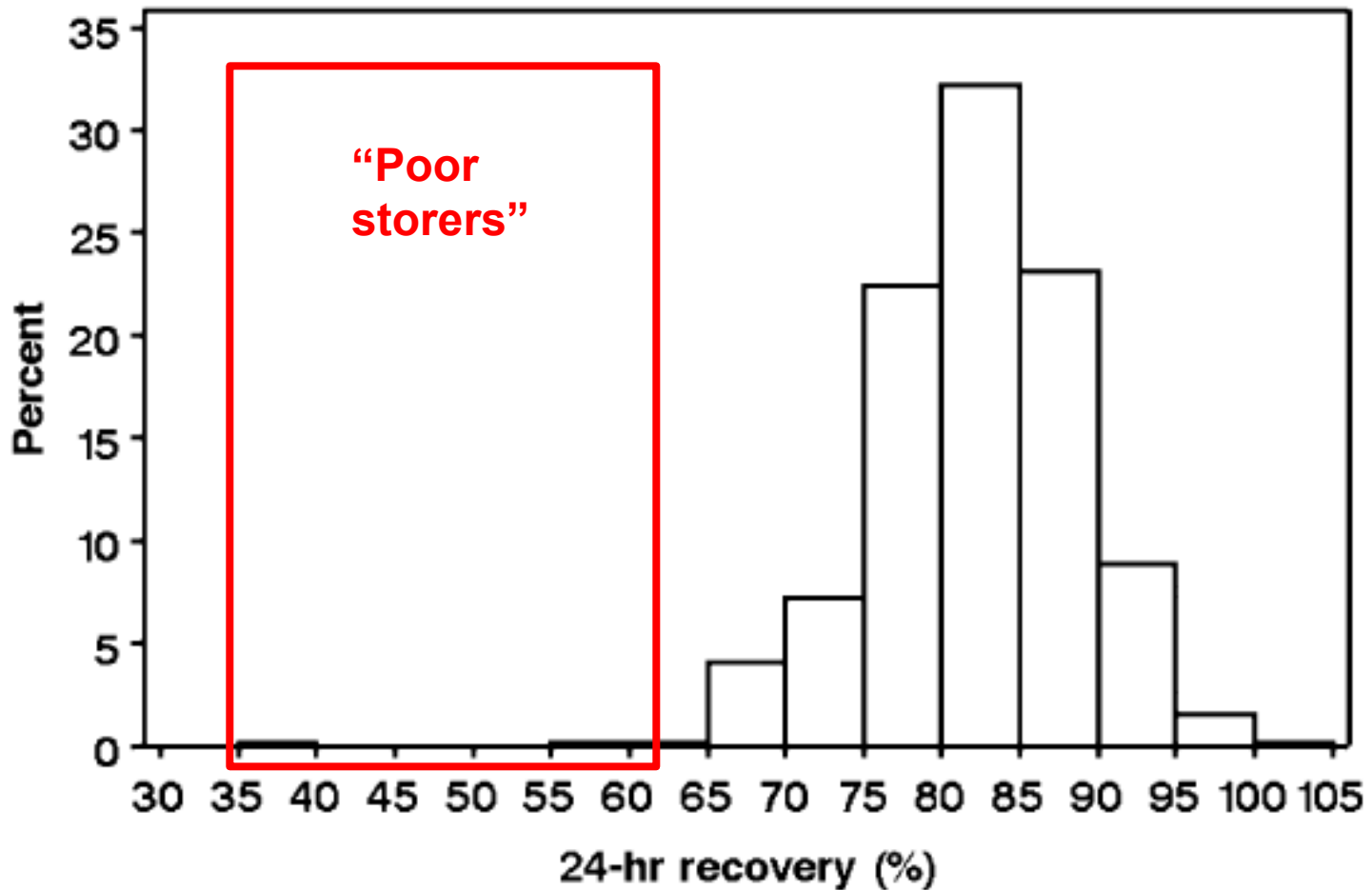


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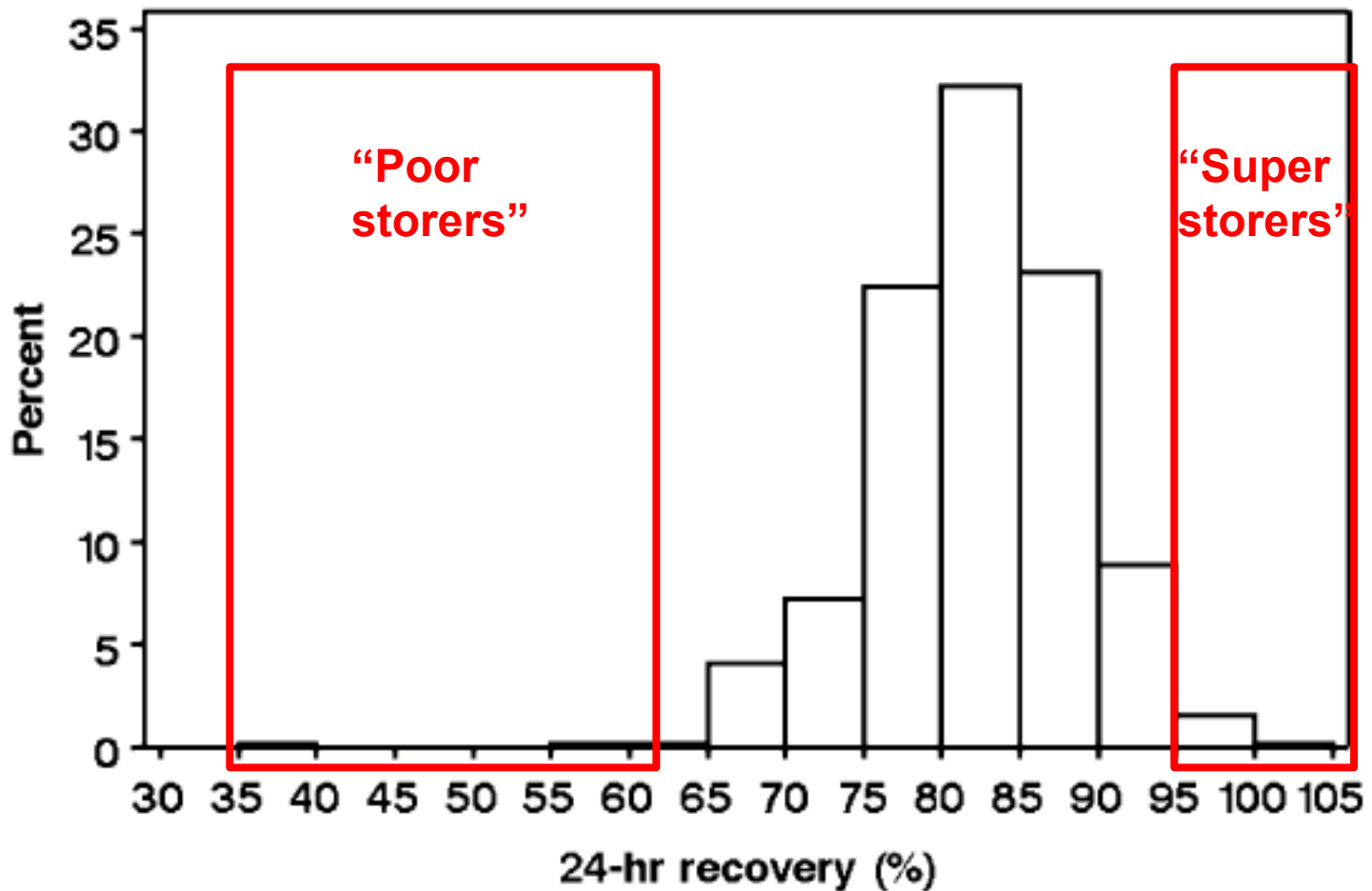
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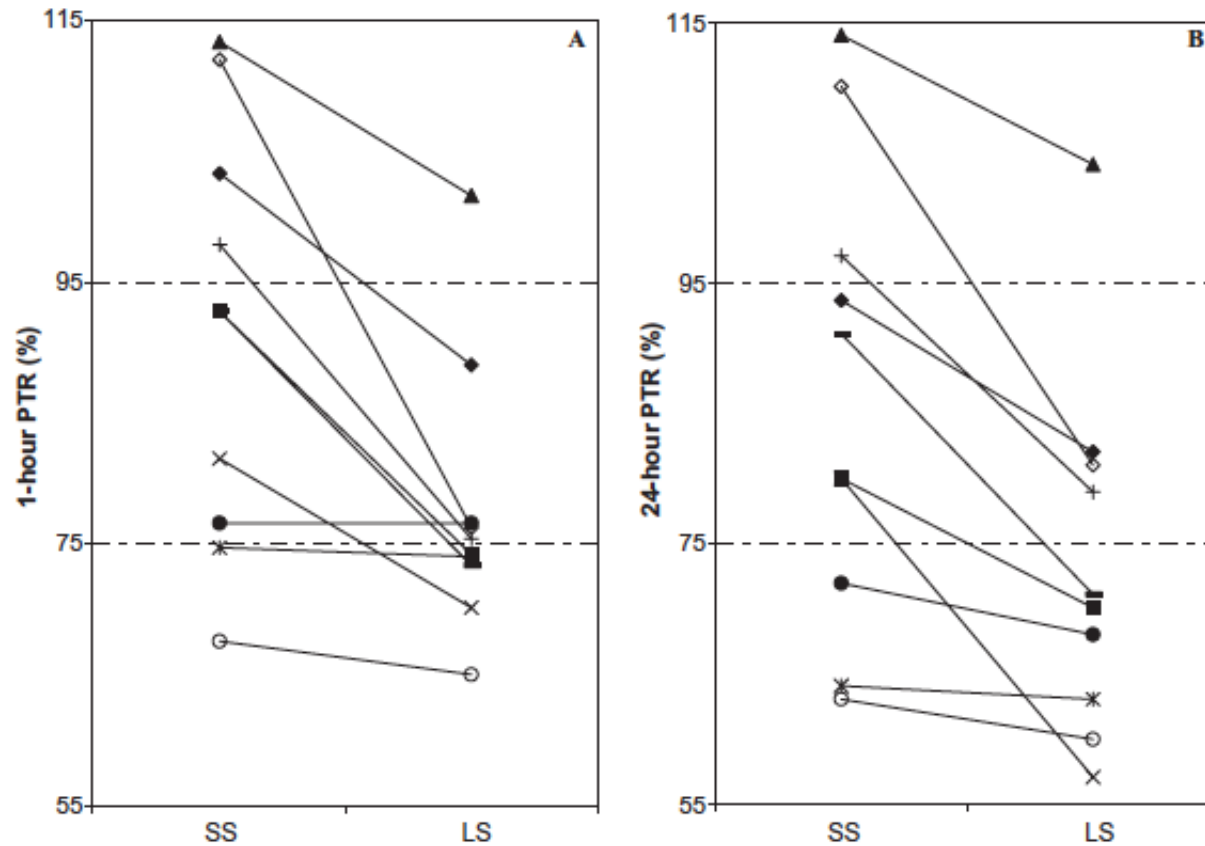


Fig. 1. Individual 1-hour PTR (A) and 24-hour PTR (B) of SS and LS RBCs. SS and LS RBCs that have been transfused into the same patient are connected to each other. Each symbol represents a patient.

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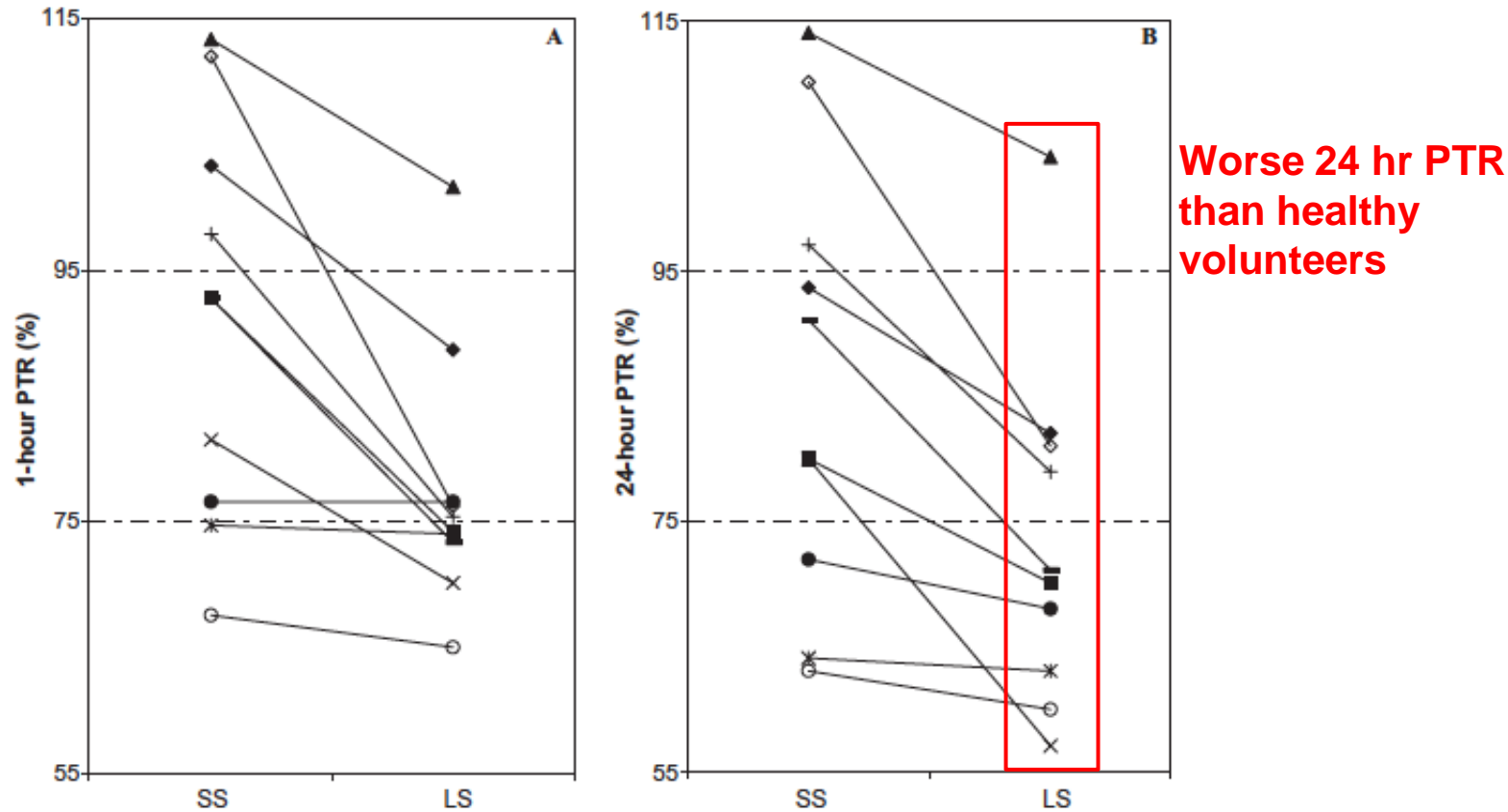


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## TRANSFUSION MEDICINE

Effect of donor, component, and recipient characteristics  
on hemoglobin increments following red blood  
cell transfusion

Nareg H. Roubinian,<sup>1-3</sup> Colleen Plimier,<sup>1</sup> Jennifer P. Woo,<sup>4</sup> Catherine Lee,<sup>1</sup> Roberta Bruhn,<sup>2,3</sup> Vincent X. Liu,<sup>1</sup> Gabriel J. Escobar,<sup>1</sup>  
Steven H. Kleinman,<sup>5</sup> Darrell J. Triulzi,<sup>6</sup> Edward L. Murphy,<sup>3,2</sup> and Michael P. Busch<sup>2,3</sup>

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# **Adverse consequences of RBC transfusion**

**Infectious  
Immunological  
Volume related  
Other (“old” RBCs?)**

**Is “old” blood bad?**

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**Infection?**

**Inflammation?**

**Thrombosis?**

**Mortality?**

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**Not going to talk about these now**

**We can discuss these later, if you  
would like**

# Is “old” blood bad?

↑ RBC refrigerated storage time



↑ RBC storage lesion *in vitro*



↓ RBC recovery *in vivo*



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**Why is transfusion of less than a full “dose” OK?**

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# **Is “old” blood bad?**

**Why is transfusion of less than a full “dose” OK?**

**Transfused RBCs that don’t circulate cannot provide their “pharmaceutical” function (e.g., O<sub>2</sub> delivery)**

**What other drug do we give whose potency decreases over time? And that’s OK?**

**Not just RBC quality**

**Not just RBC quality**

**Activity of recipient's  
mononuclear phagocyte system**

**(1) Quality of transfused donor RBCs**

**and**

**(2) activity of recipient mononuclear  
phagocyte system**

**determine post-transfusion recovery  
and RBC lifespan *in vivo***

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**When do RBCs “go bad”?**



# When do RBCs “go bad”?

The Journal of Clinical Investigation

CLINICAL MEDICINE

## Prolonged red cell storage before transfusion increases extravascular hemolysis

Francesca Rapido,<sup>1,2</sup> Gary M. Brittenham,<sup>3,4</sup> Sheila Bandyopadhyay,<sup>1</sup> Francesca La Carpia,<sup>1</sup> Camilla L'Acqua,<sup>1</sup> Donald J. McMahon,<sup>4</sup> Abdelhadi Rebbaa,<sup>1</sup> Boguslaw S. Wojczyk,<sup>1</sup> Jane Netterwald,<sup>1</sup> Hangli Wang,<sup>1</sup> Joseph Schwartz,<sup>1</sup> Andrew Eisenberger,<sup>4</sup> Mark Soffing,<sup>5</sup> Randy Yeh,<sup>5</sup> Chaitanya Divgi,<sup>5</sup> Yelena Z. Ginzburg,<sup>6</sup> Beth H. Shaz,<sup>6</sup> Sujit Sheth,<sup>7</sup> Richard O. Francis,<sup>1</sup> Steven L. Spitalnik,<sup>1</sup> and Eldad A. Hod<sup>1</sup>

Journal of Clinical Investigation 127:375-382, 2017



**E. Hod**

**60 healthy volunteers enrolled**

**52 completed study**

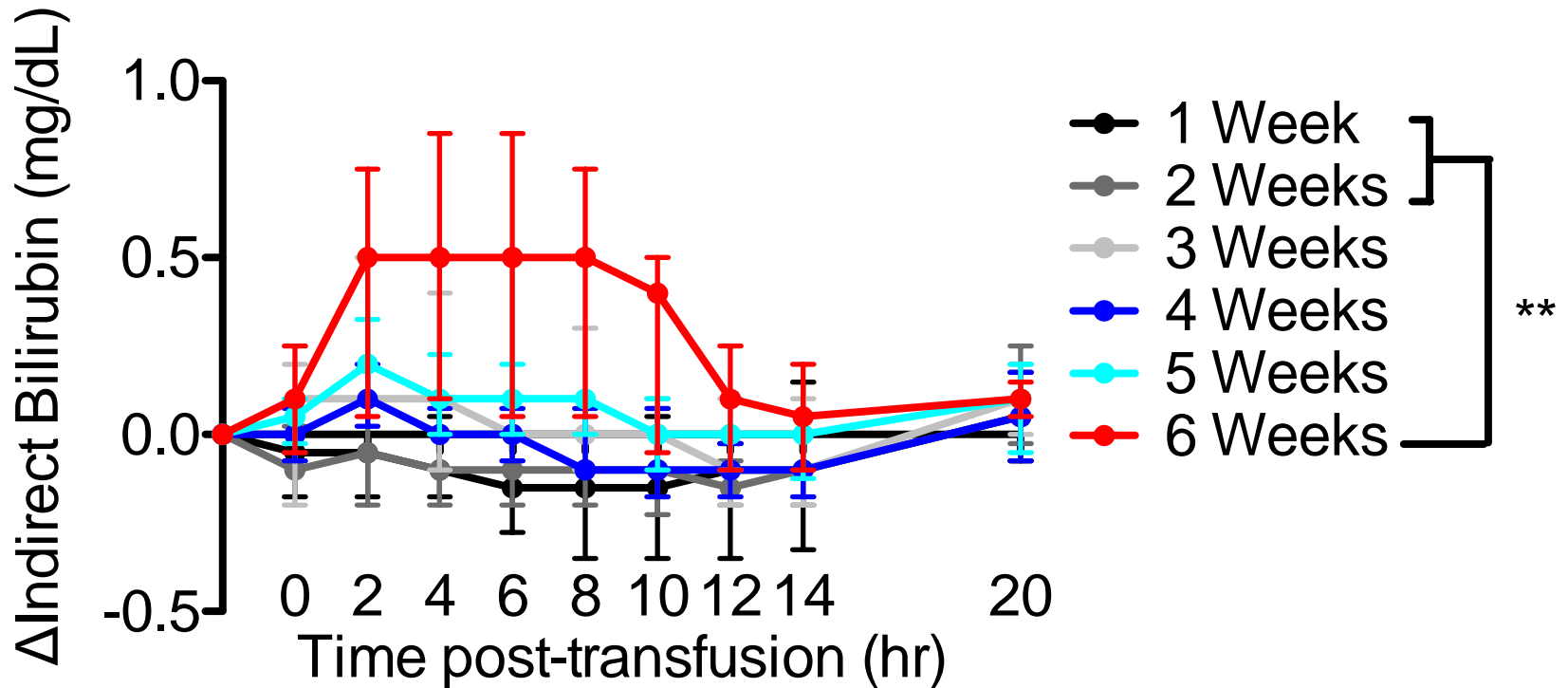
**Randomized to 1, 2, 3, 4, 5, 6 weeks of storage**

**Donation at NYBC; leukoreduced; AS-3**

**Transfused with entire unit**

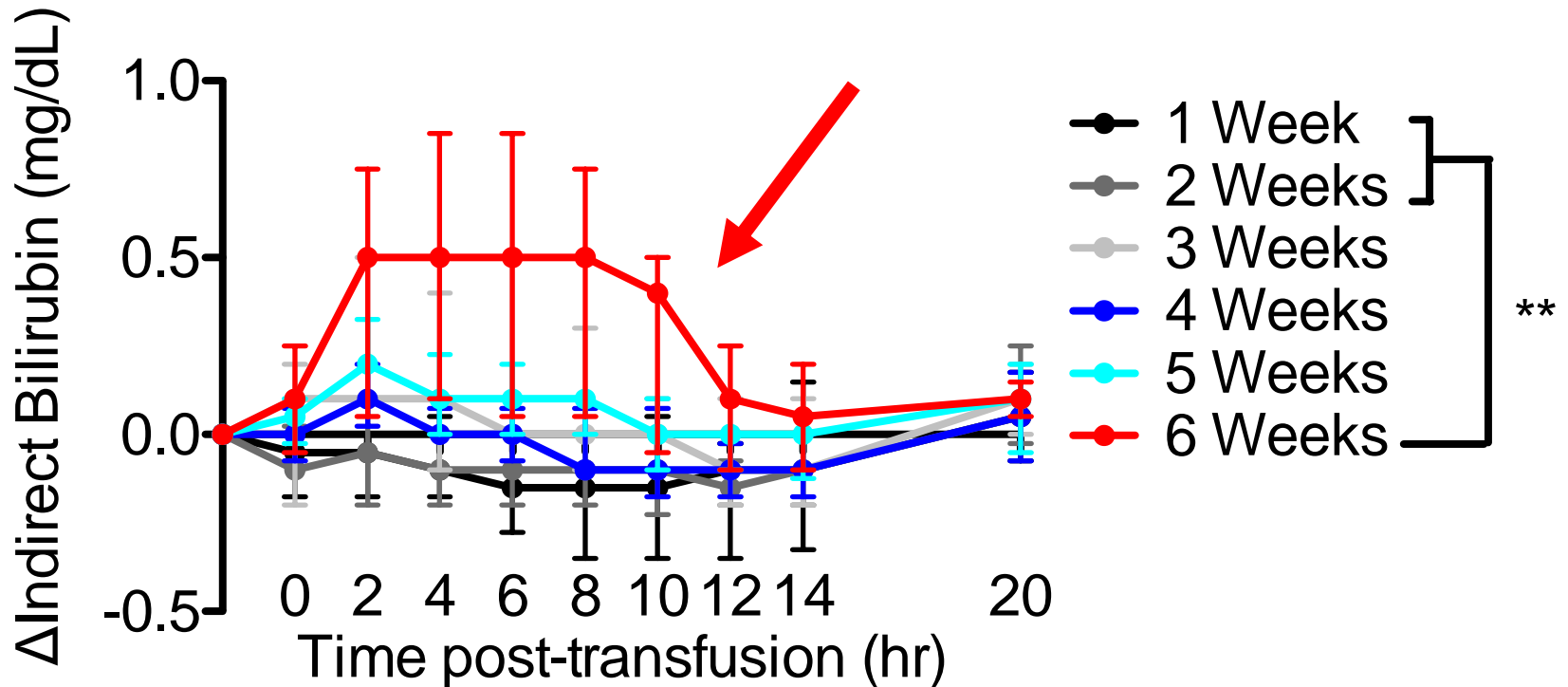
**$^{51}\text{Cr}$ -labeled post-transfusion recovery**

# When do RBCs “go bad”?



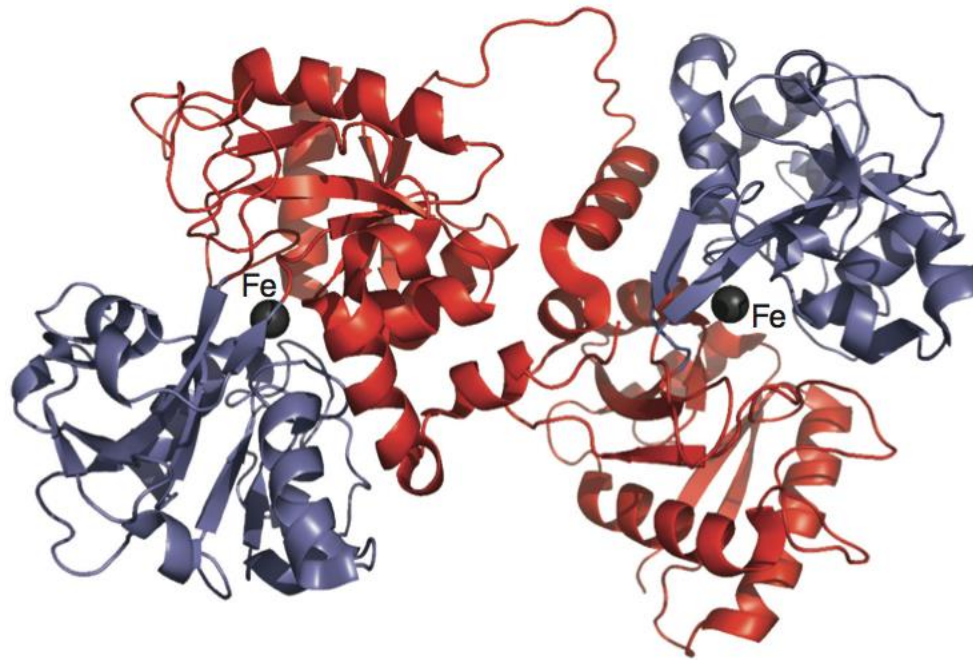
# When do RBCs “go bad”?

**6 Weeks**



# Transferrin Saturation ( $Tf_{sat}$ )

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<http://www.chemtube3d.com/solidstate/BC-26-13.htm>

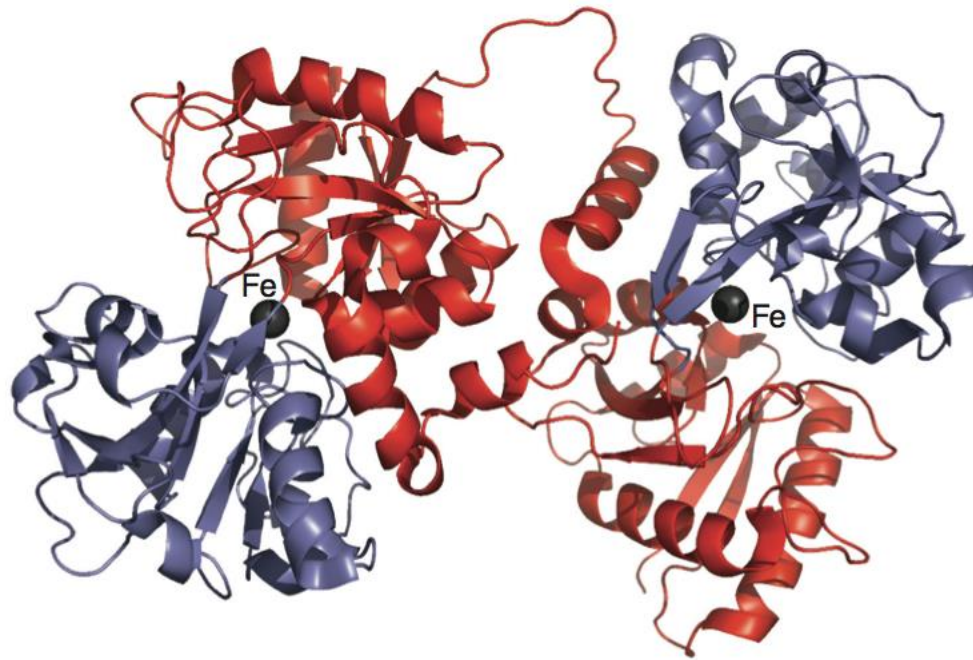
**Apo-Tf: no iron bound**

**Holo-Tf: 2 iron atoms bound**

**$Tf_{sat} = 20\% \rightarrow 80\%$  of iron-binding sites available**

**$Tf_{sat} = 75-80\% \rightarrow$  NTBI appears**

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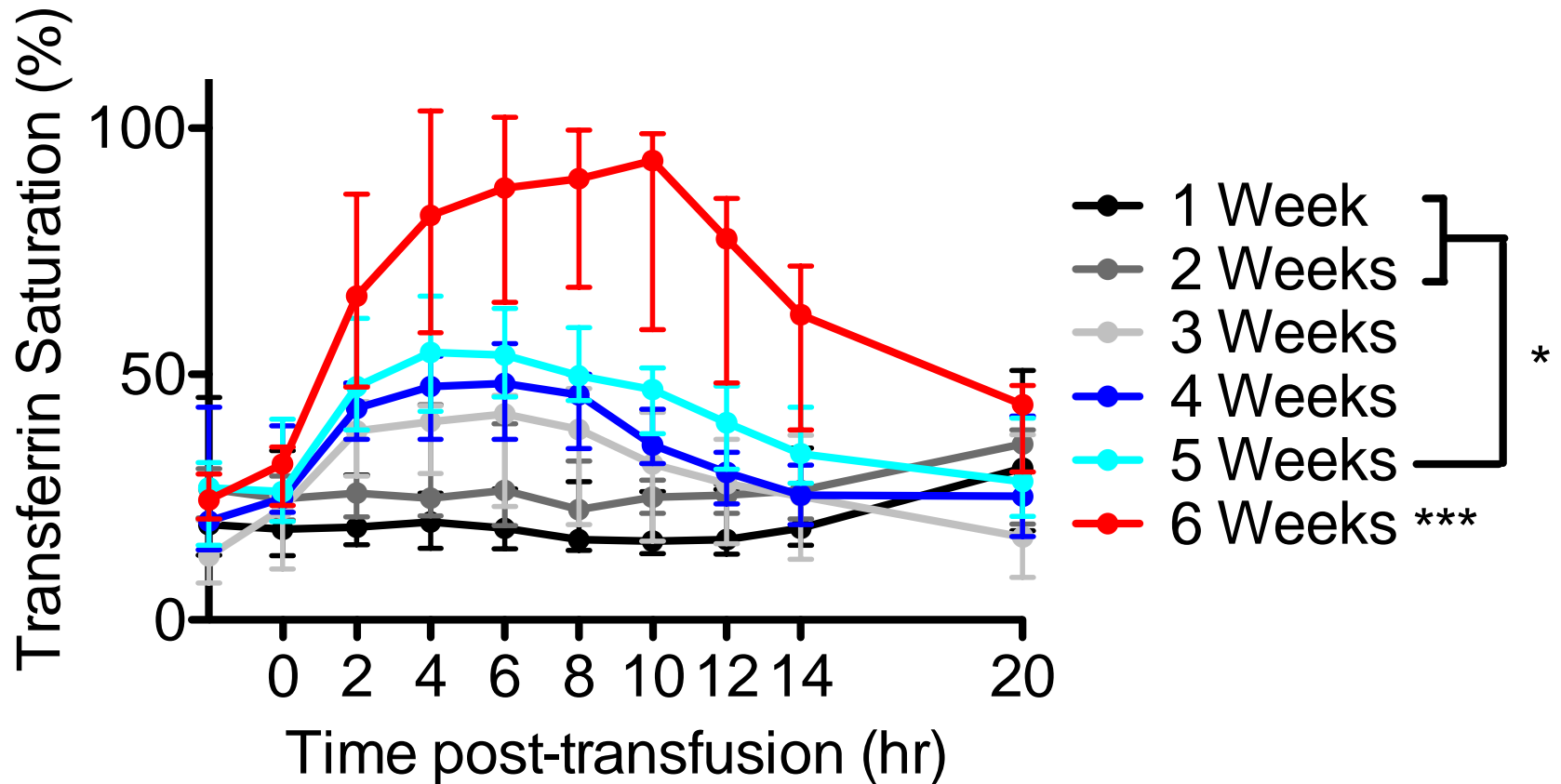
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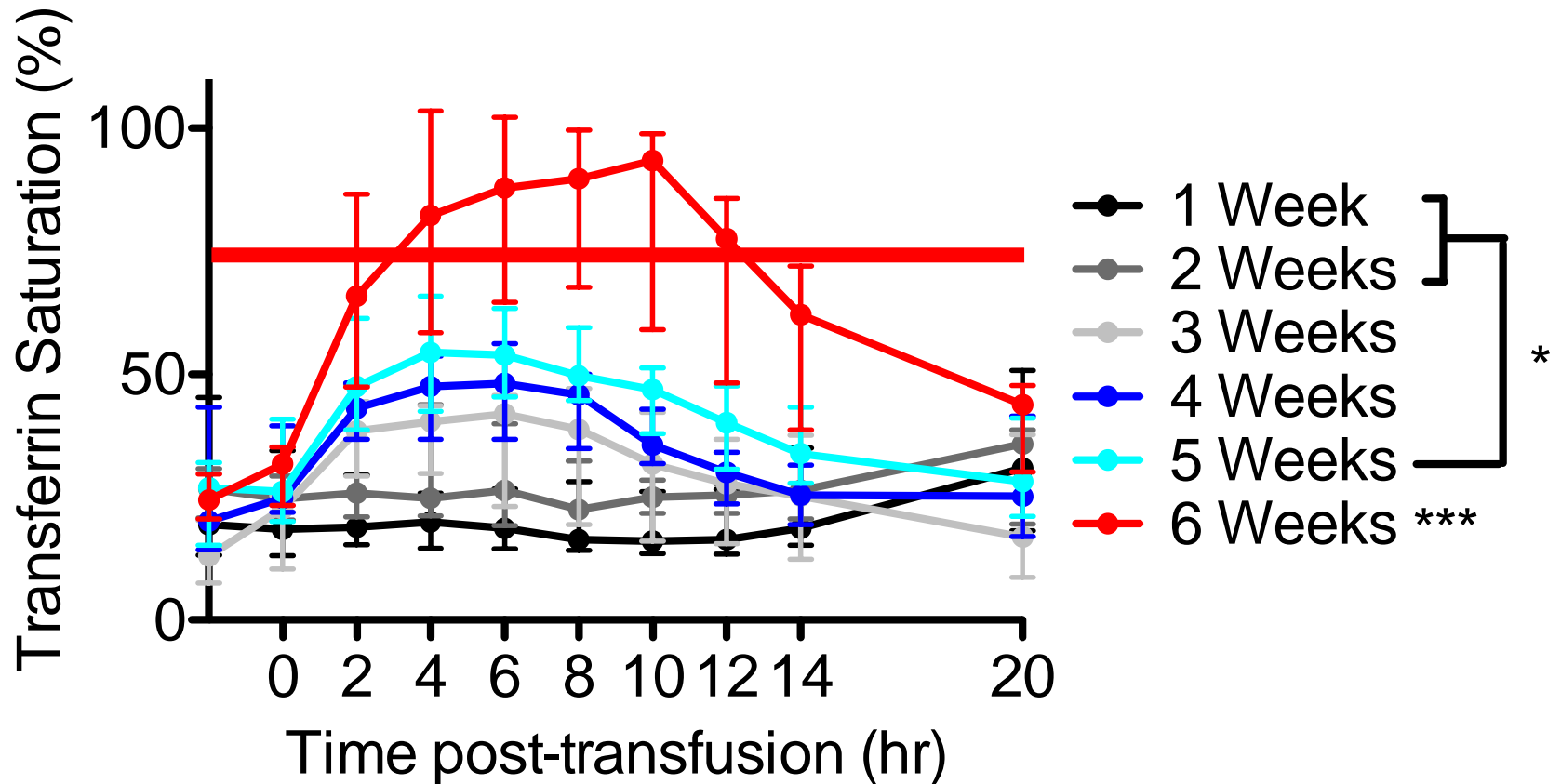
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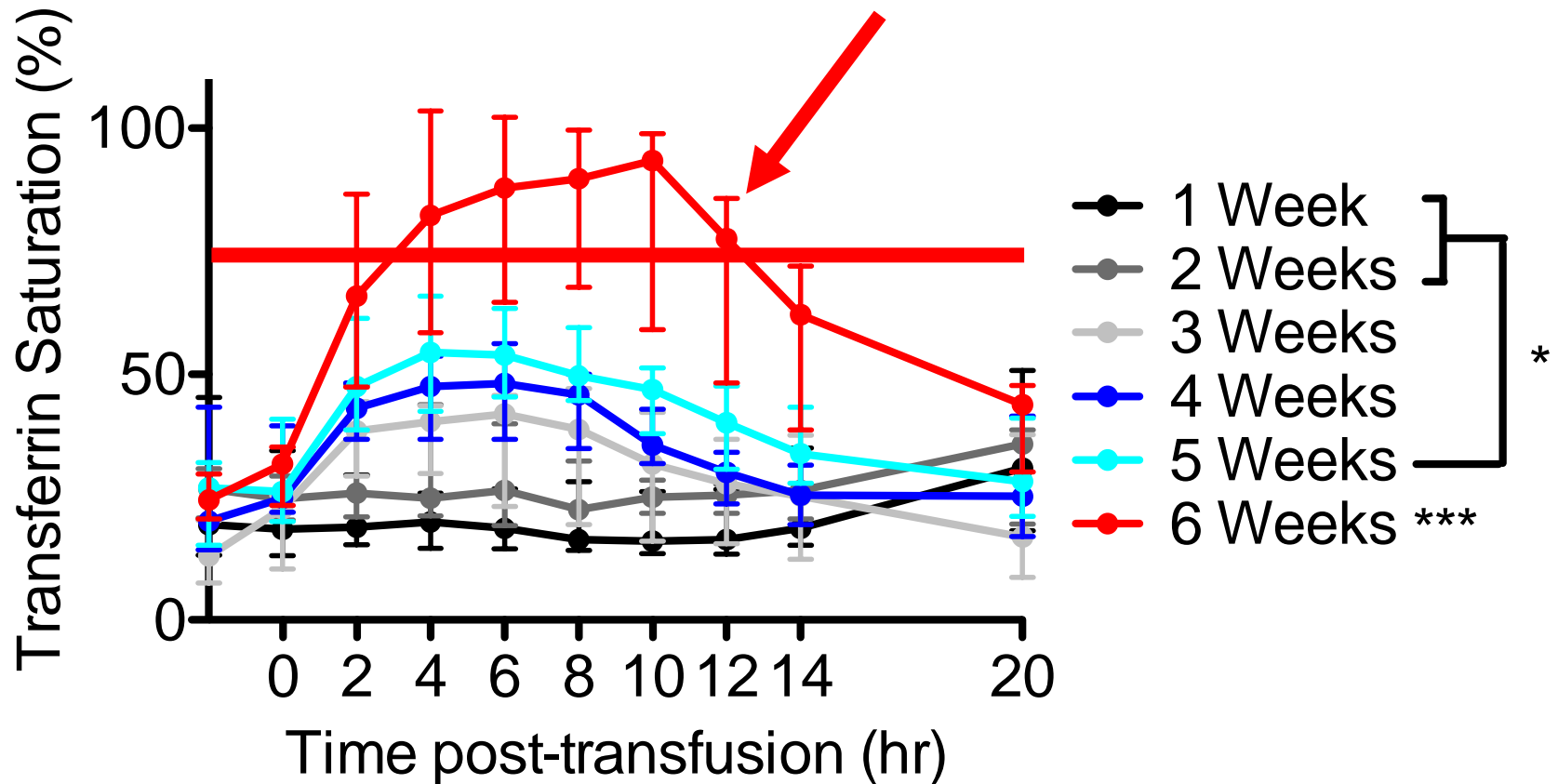


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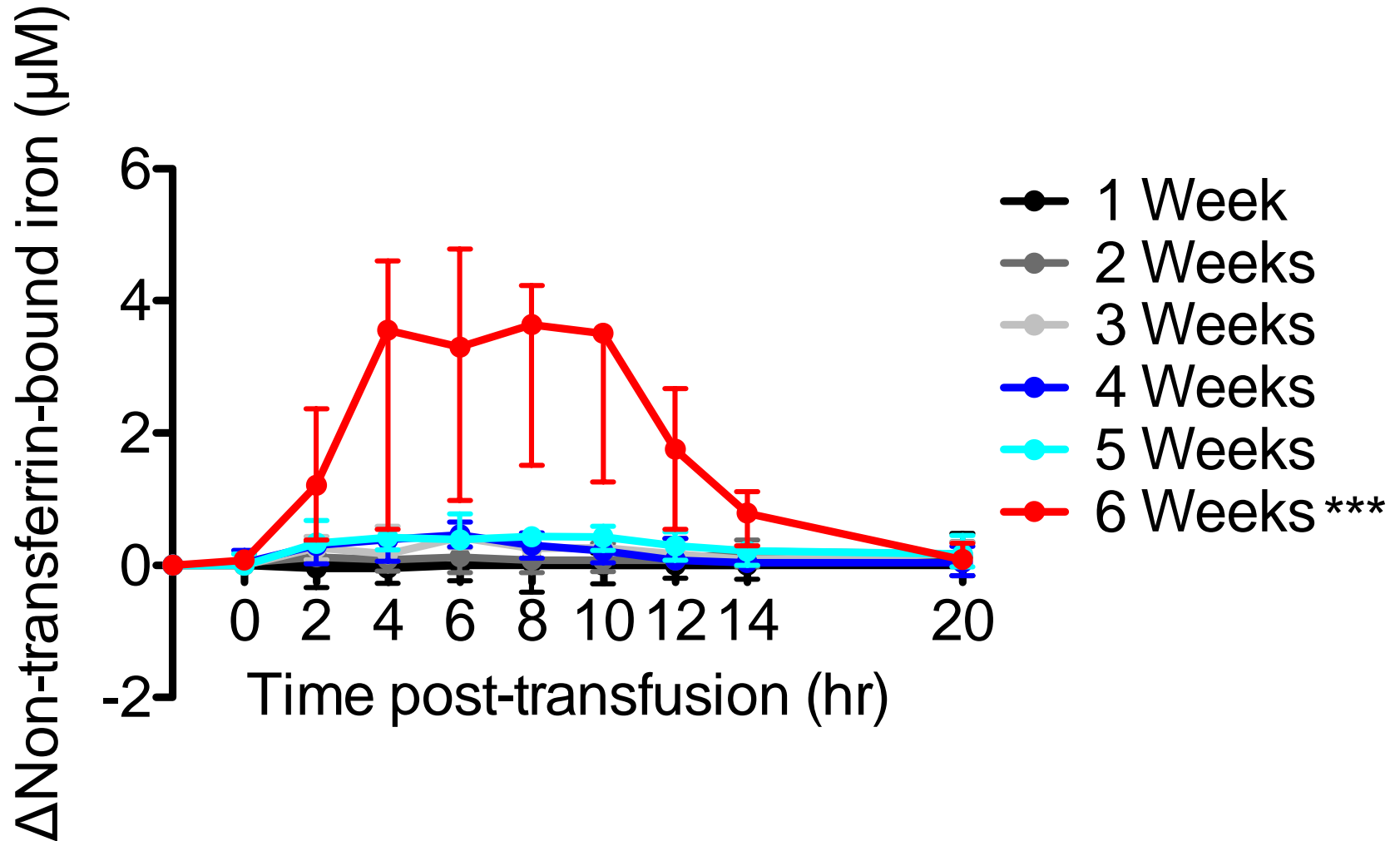


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**6 Weeks**

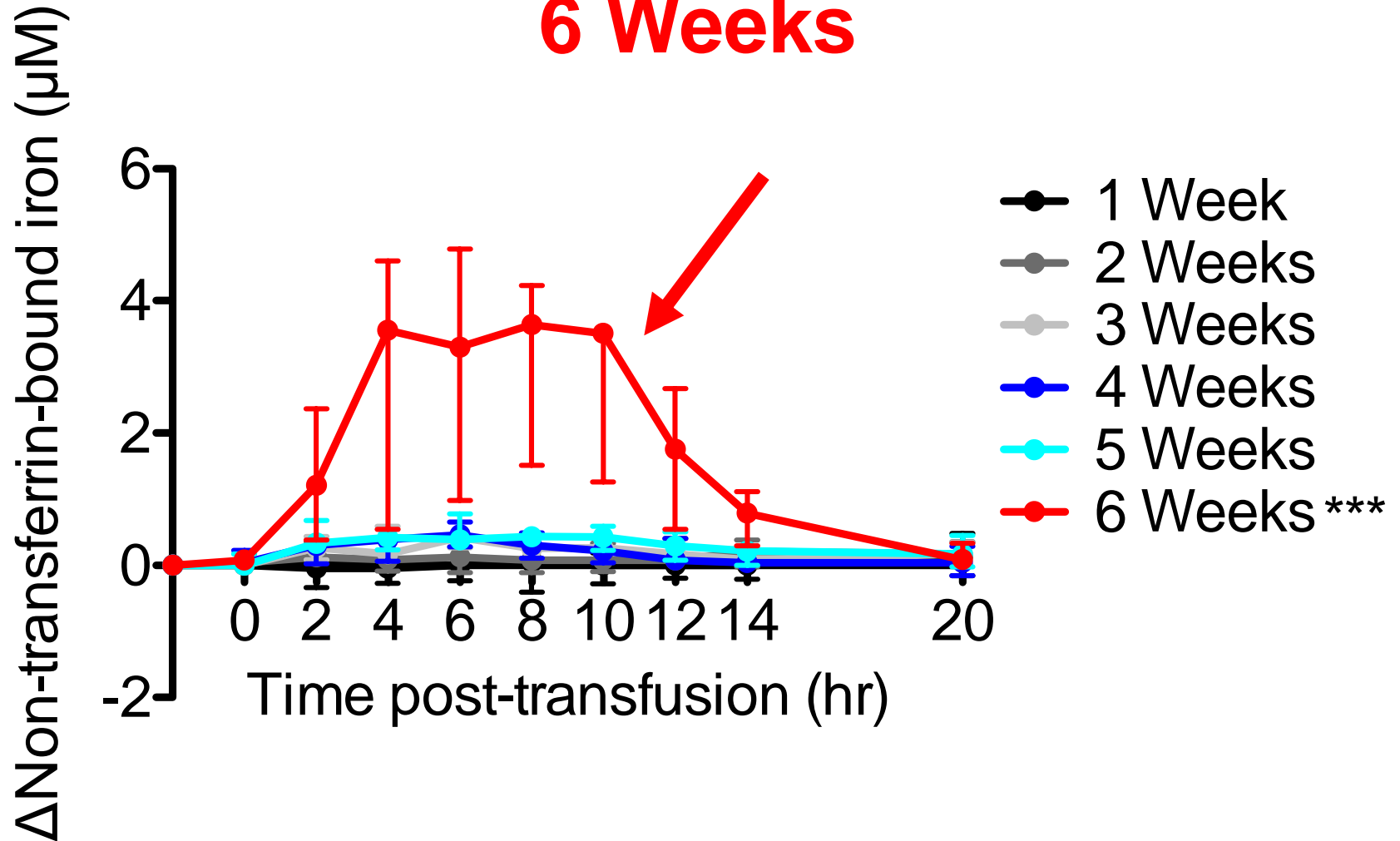


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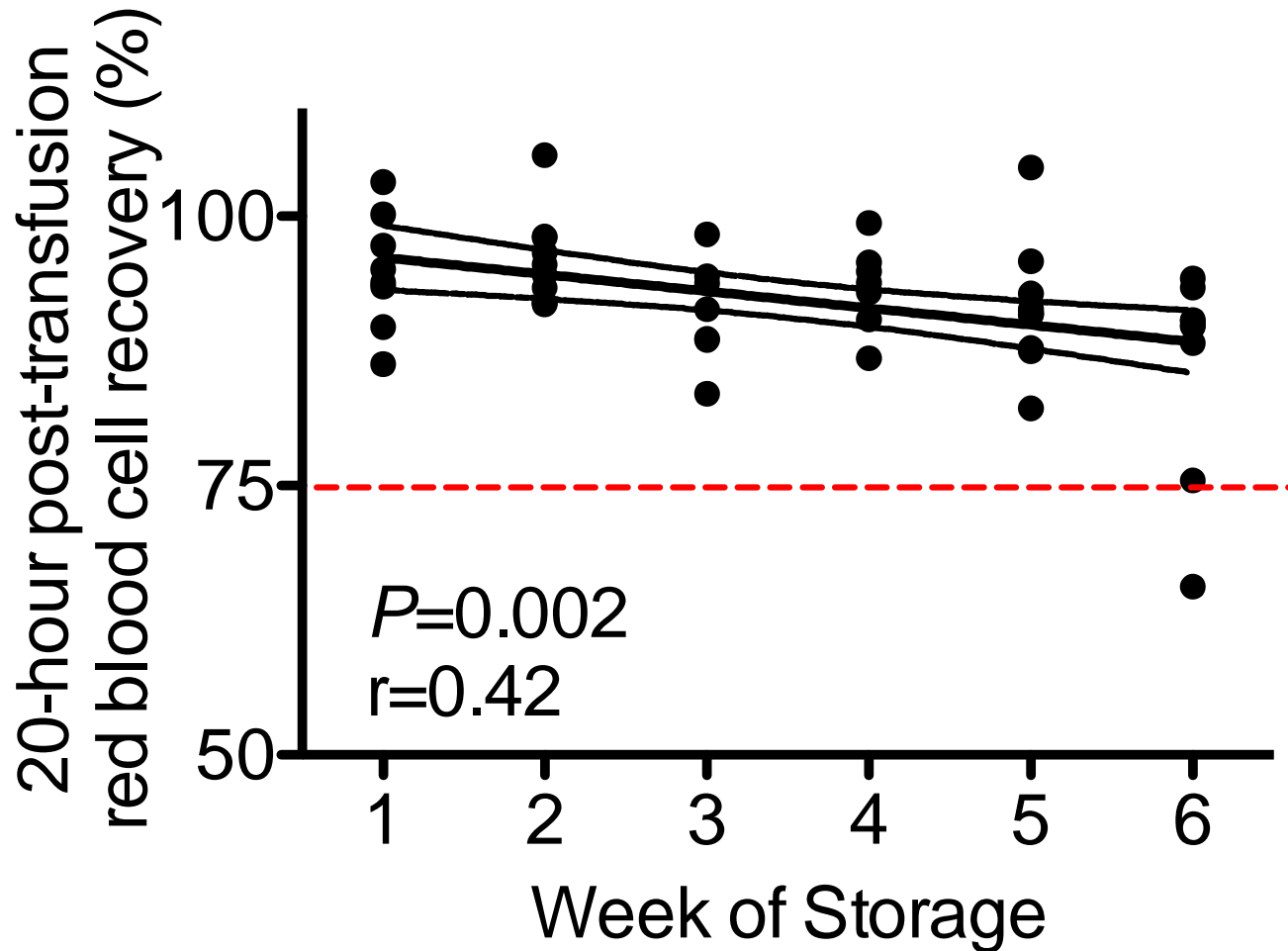


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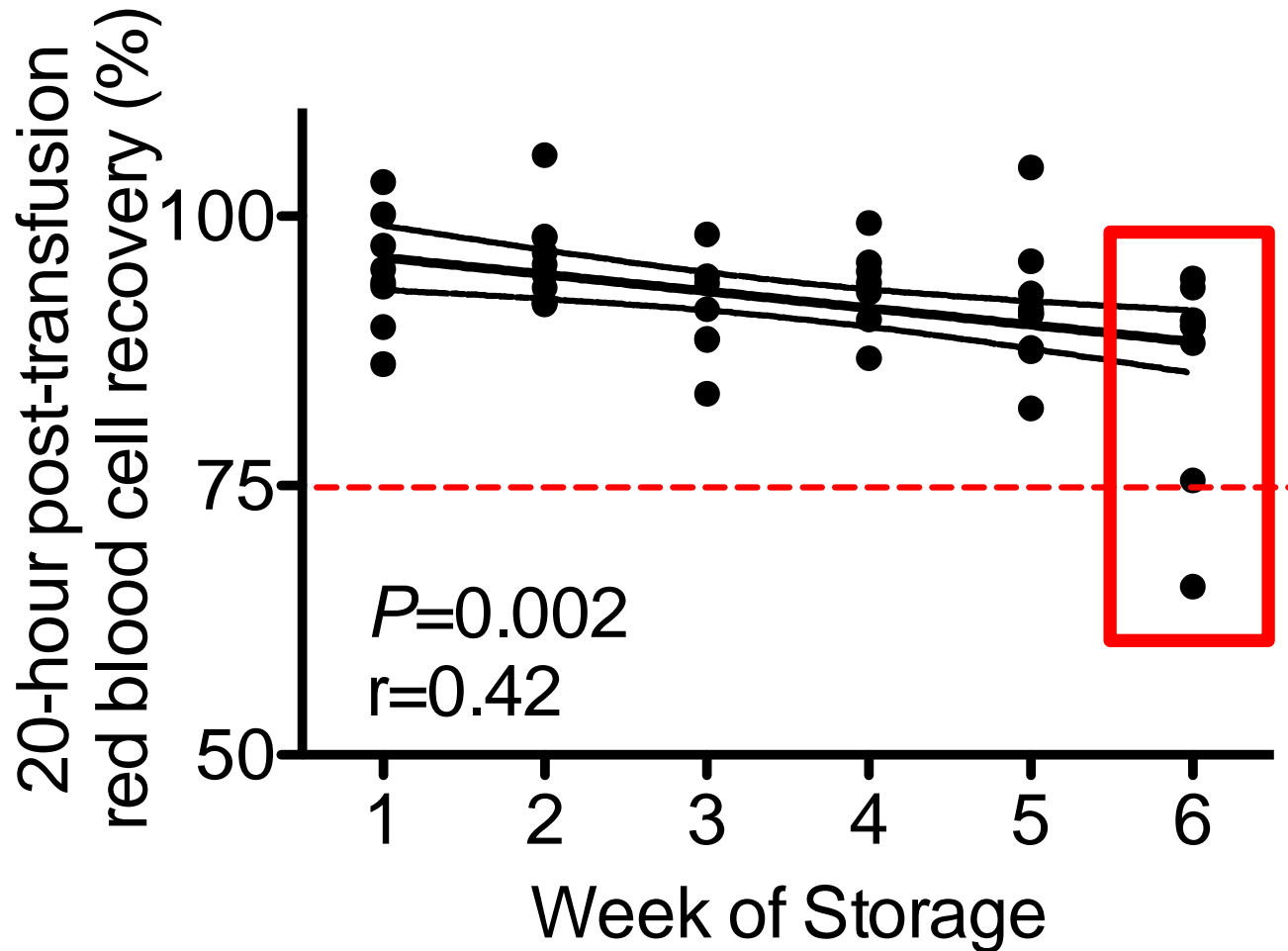
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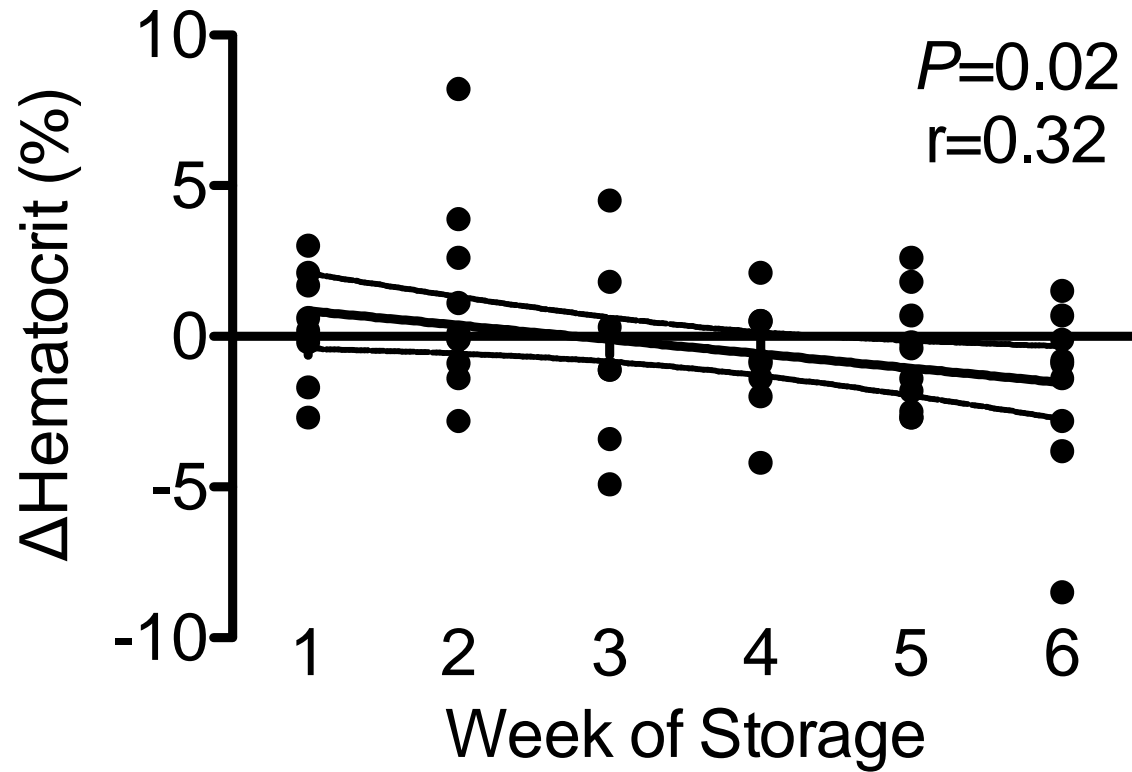
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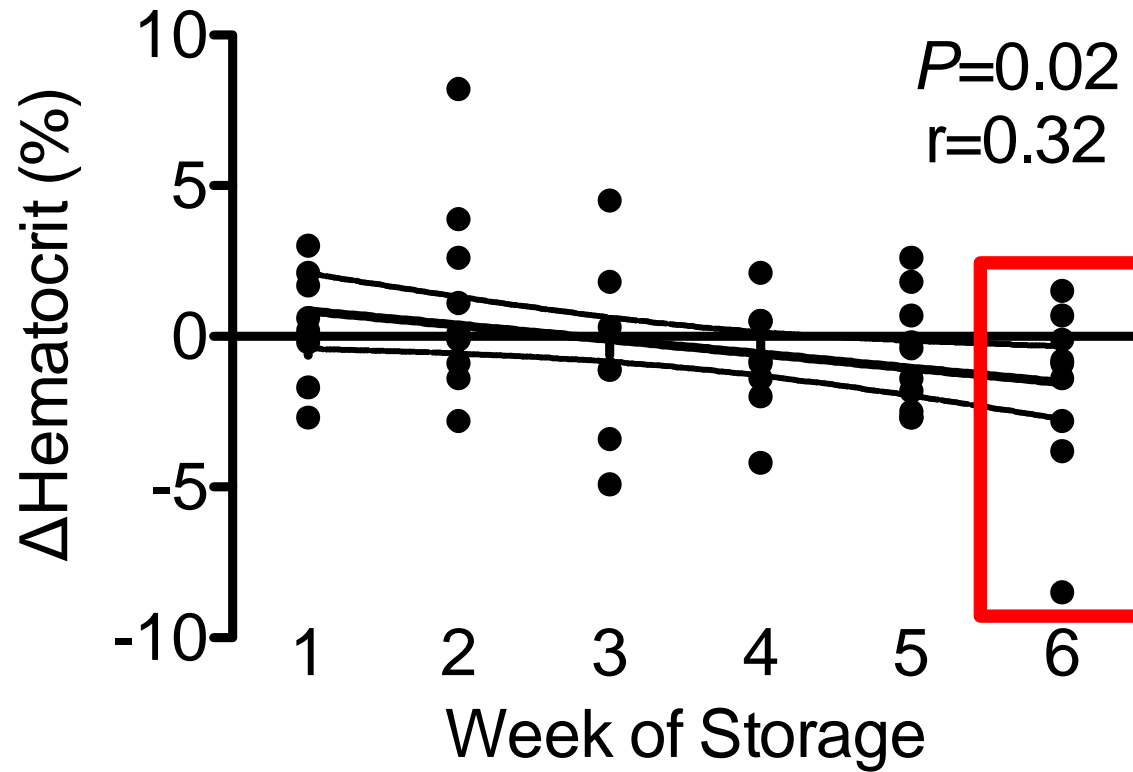
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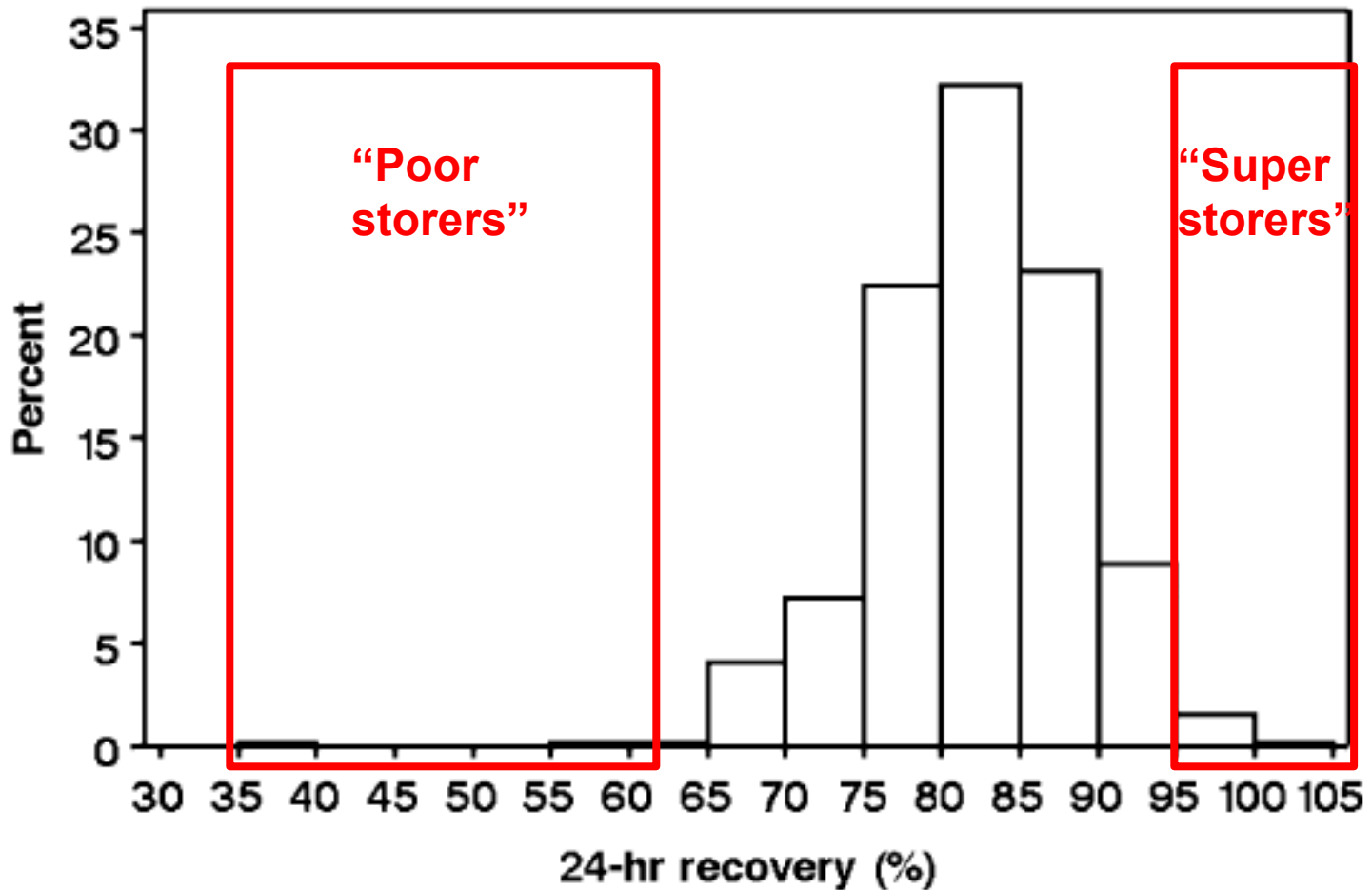




**What influences variation in  
post-transfusion recovery?**

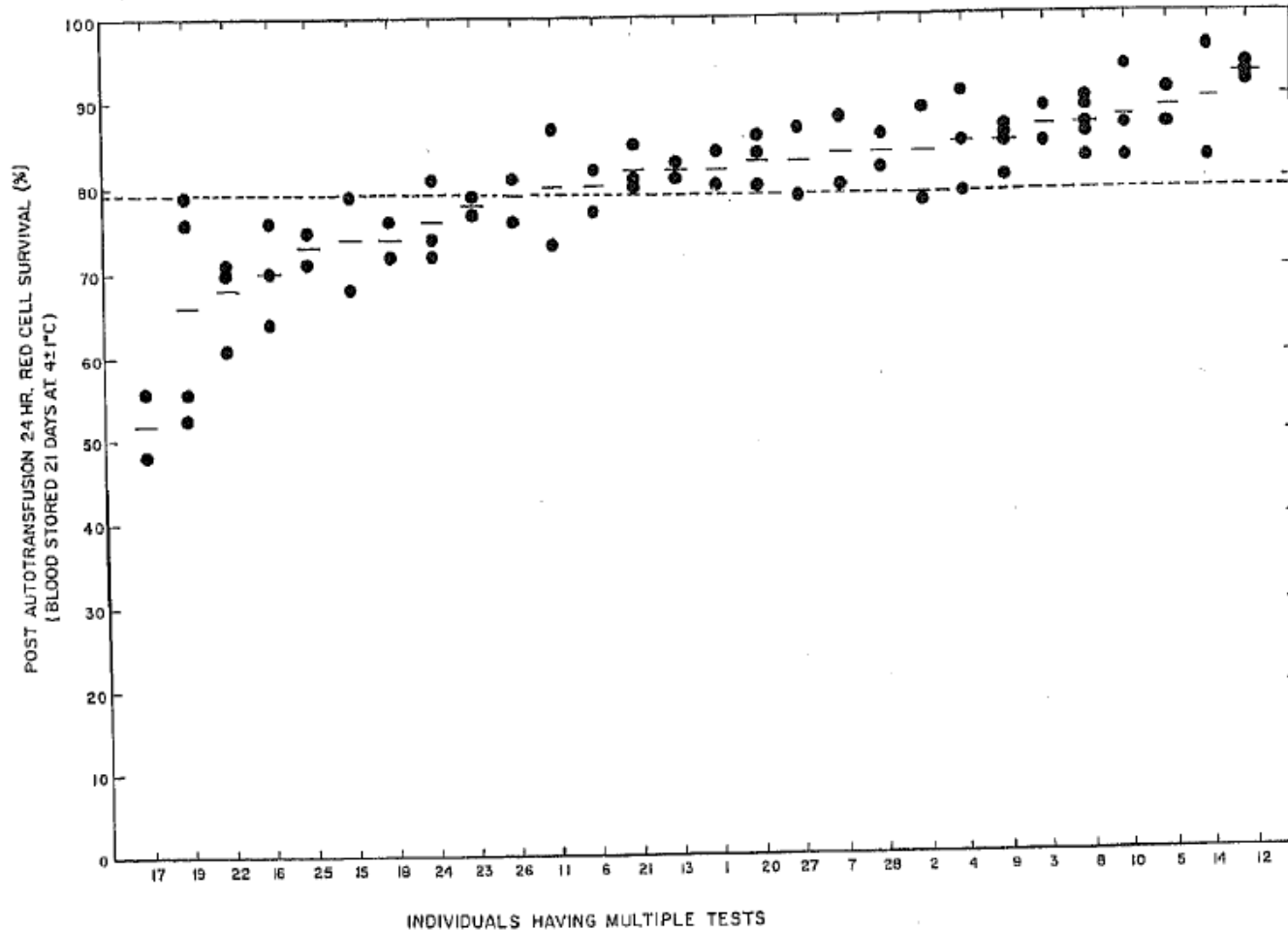
# Genetics

# 24-hr RBC recovery in 641 healthy volunteers



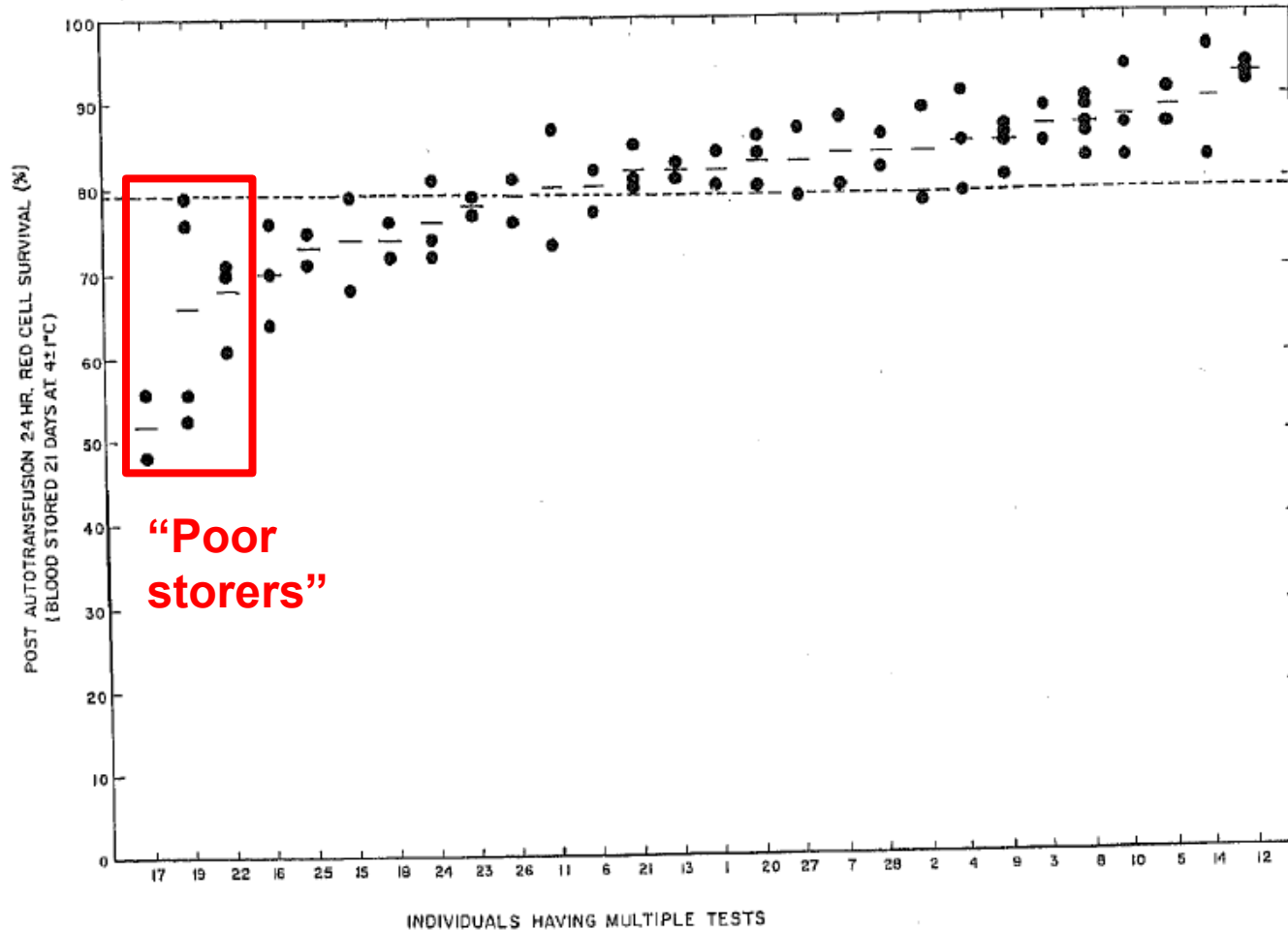
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# Repetitive 24-hr RBC recovery in 28 healthy volunteers



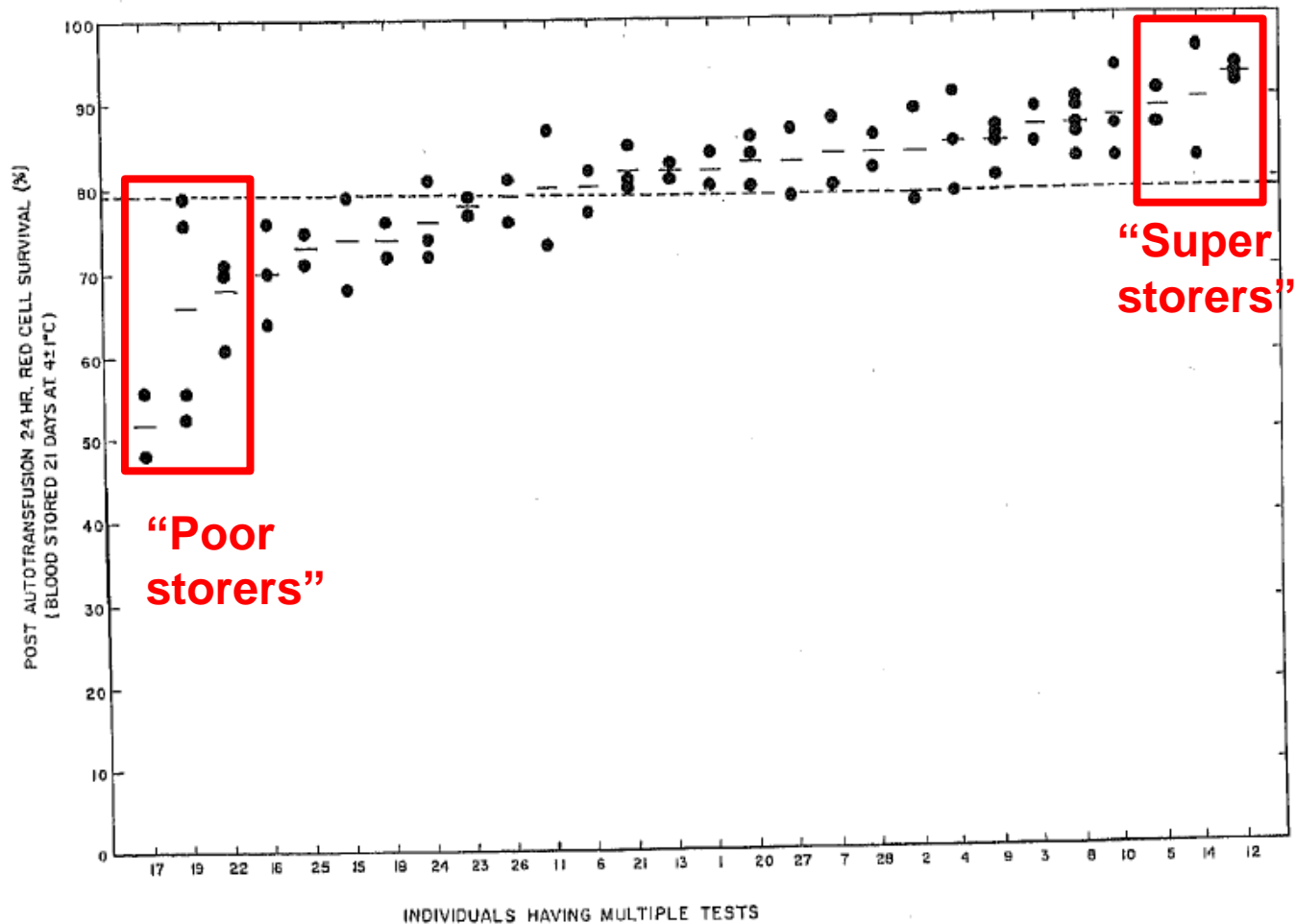
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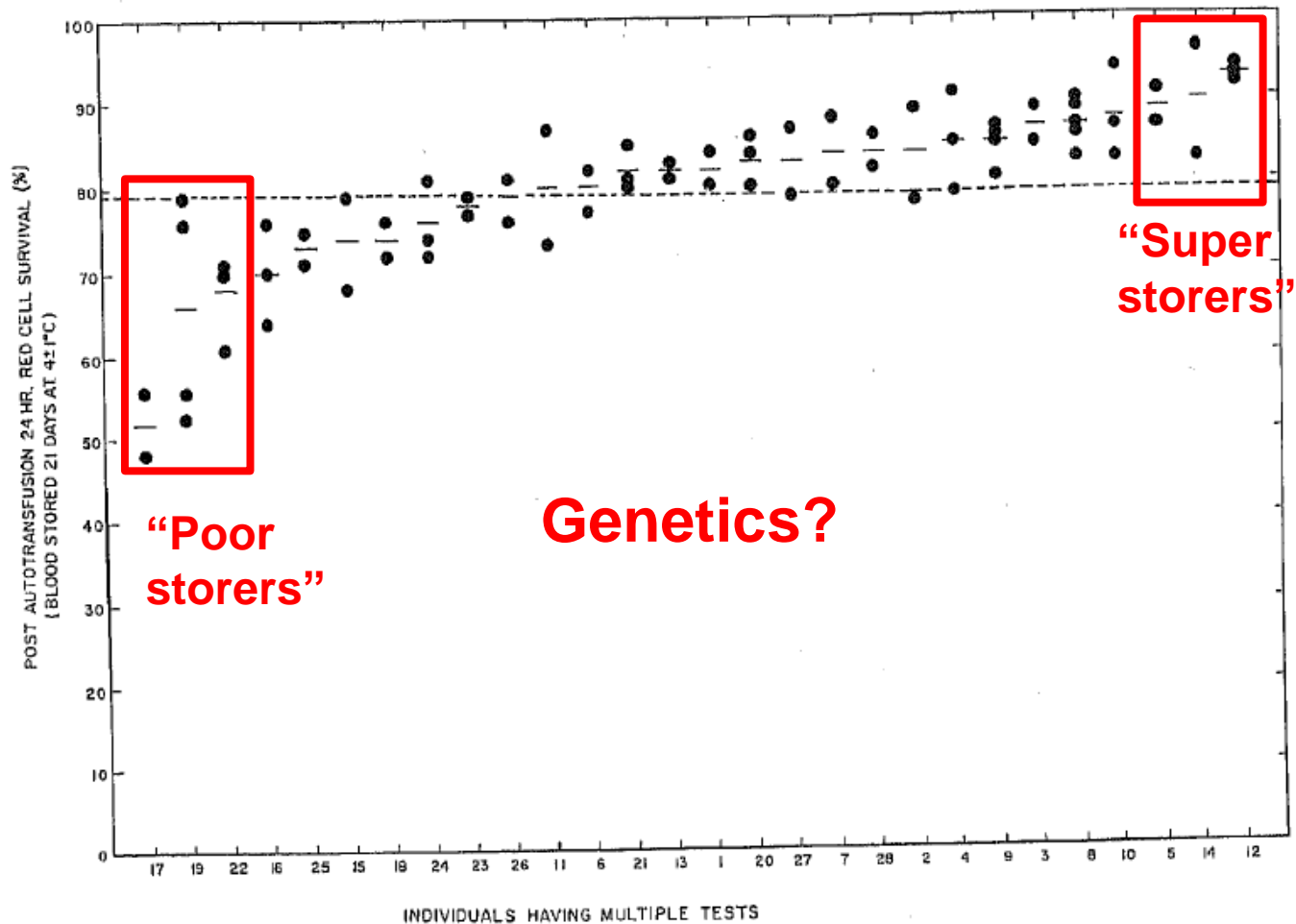
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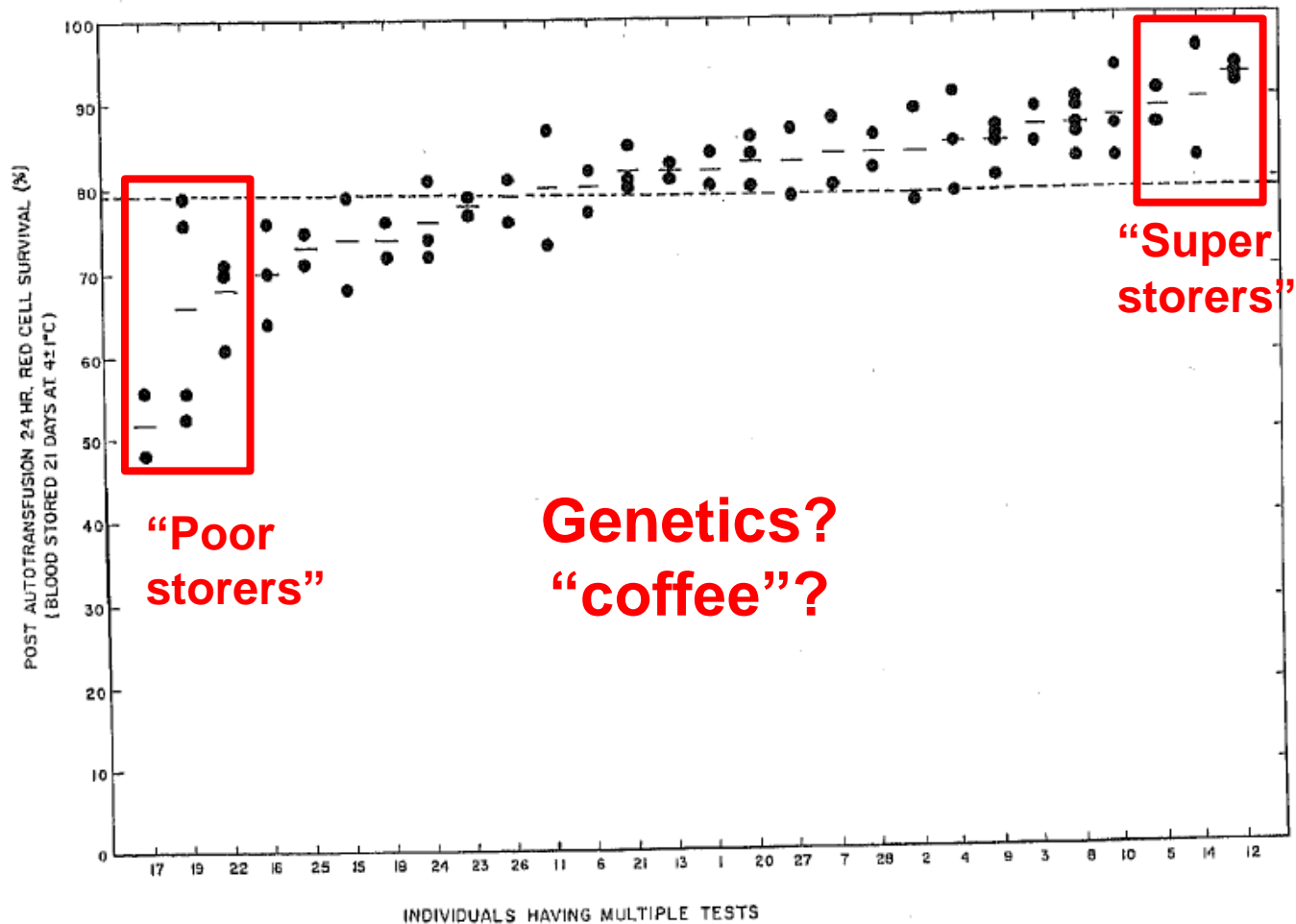
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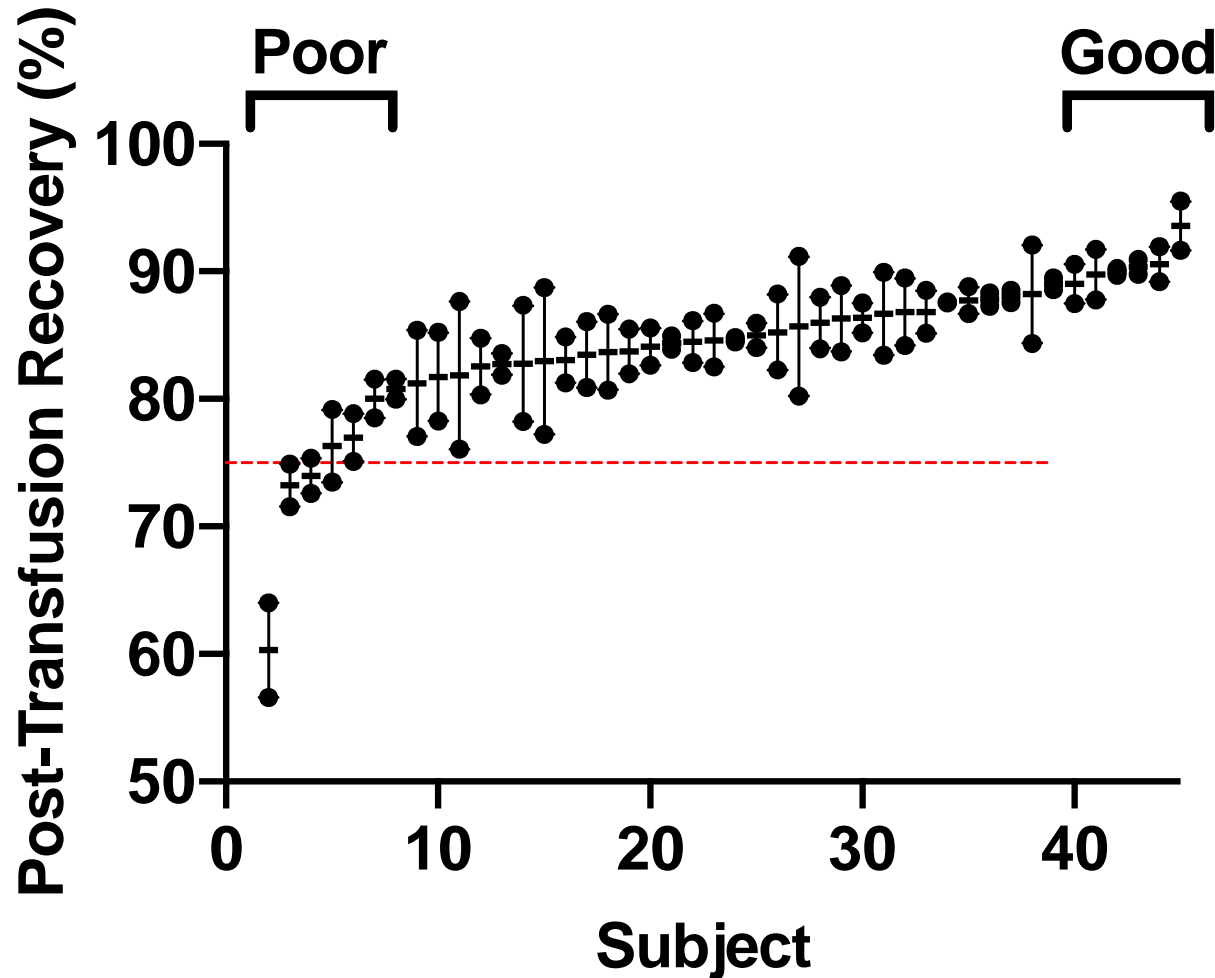
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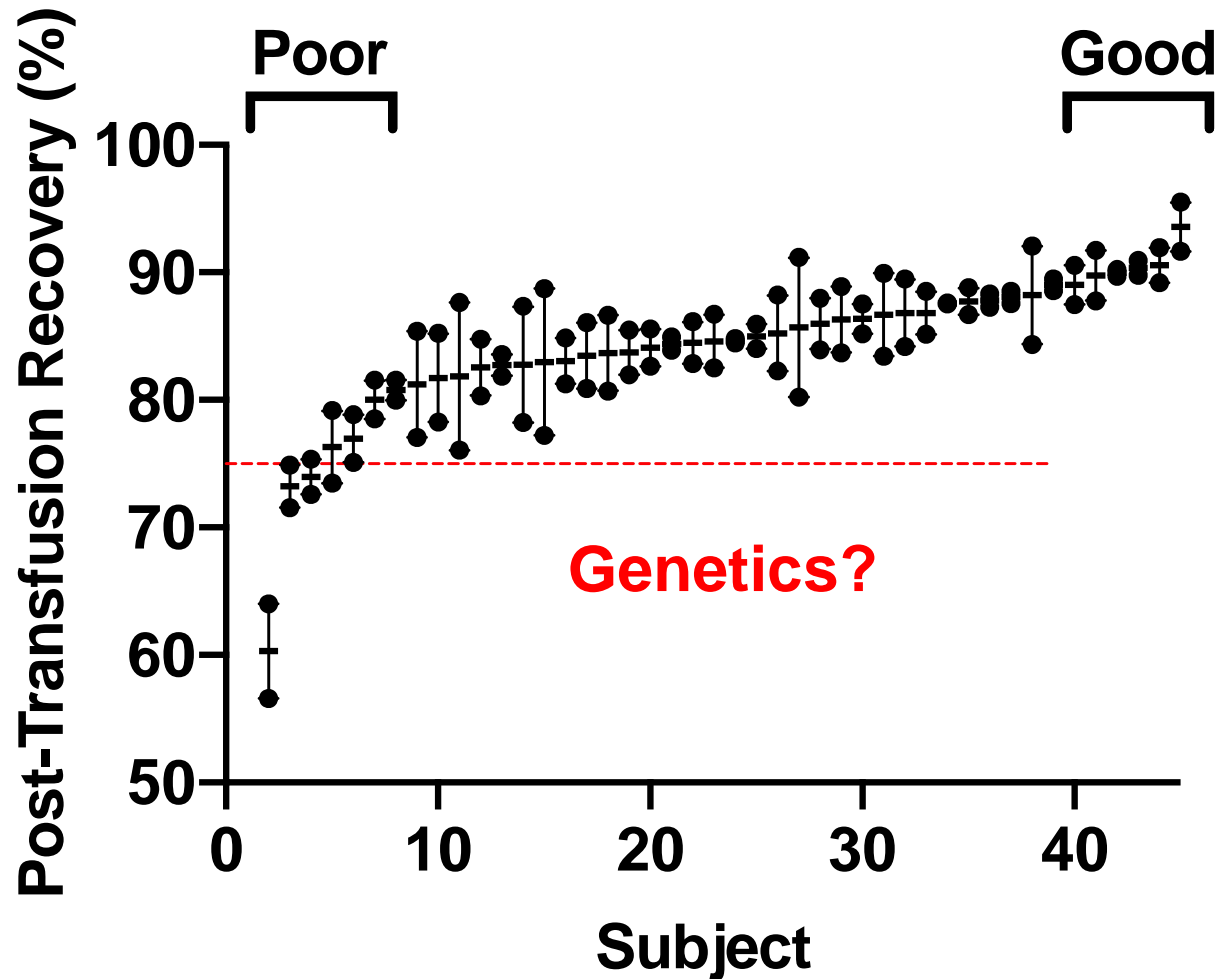
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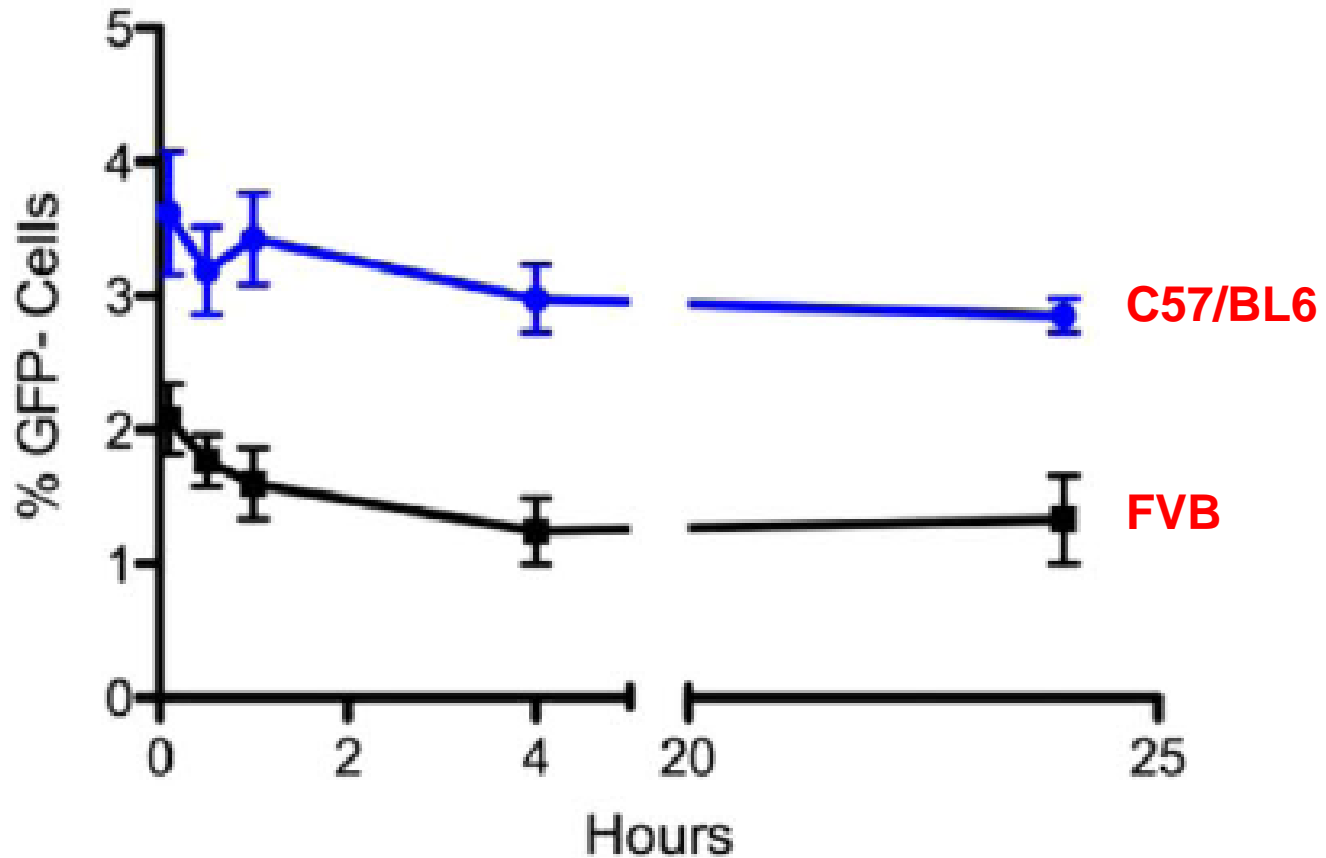
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# Repetitive 24-hr RBC recovery in 28 healthy volunteers



# 24-hr RBC recovery in inbred mouse strains



Zimring et al. Transfusion 54:137-148, 2014.

**What specific genes could be involved in RBC storage quality?**

# **General**

**Humans:**

**Gender**

**Race**

**Mice:**

**Gender**

**Strain**

# The “RBC storage lesion”

## Final common pathway?

Metabolic dysfunction & oxidative stress →

↓ Deformability

↑ “Eat me” signals

↓ “Don’t eat me” signals

↑ Hemolysis *in vitro*

↑ RBC clearance *in vivo*

Intravascular and extravascular hemolysis

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**G6PD-deficiency**

**Hemoglobin S, C, E, F, etc.**

**$\alpha$ - and  $\beta$ -thalassemias**

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**What are the consequences (if any) of the clearance of stored RBCs?**

**RBC storage lesion *in vitro***



**Decreased RBC recovery *in vivo***

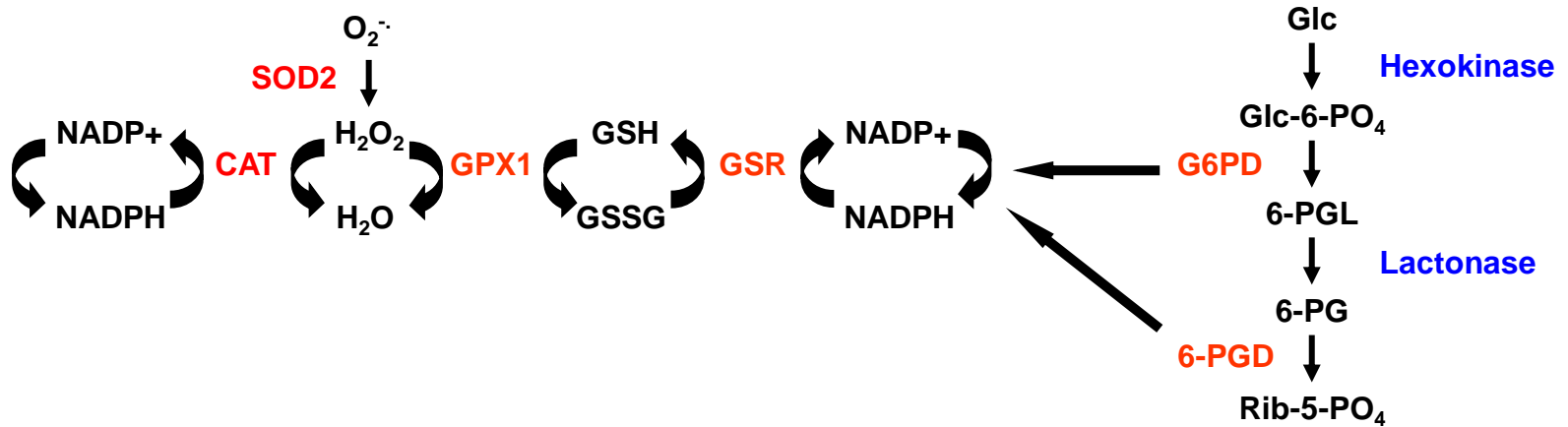
**What are the consequences (if any) of the clearance of stored RBCs?**

**Insufficient protection against  
oxidative stress *in vitro***

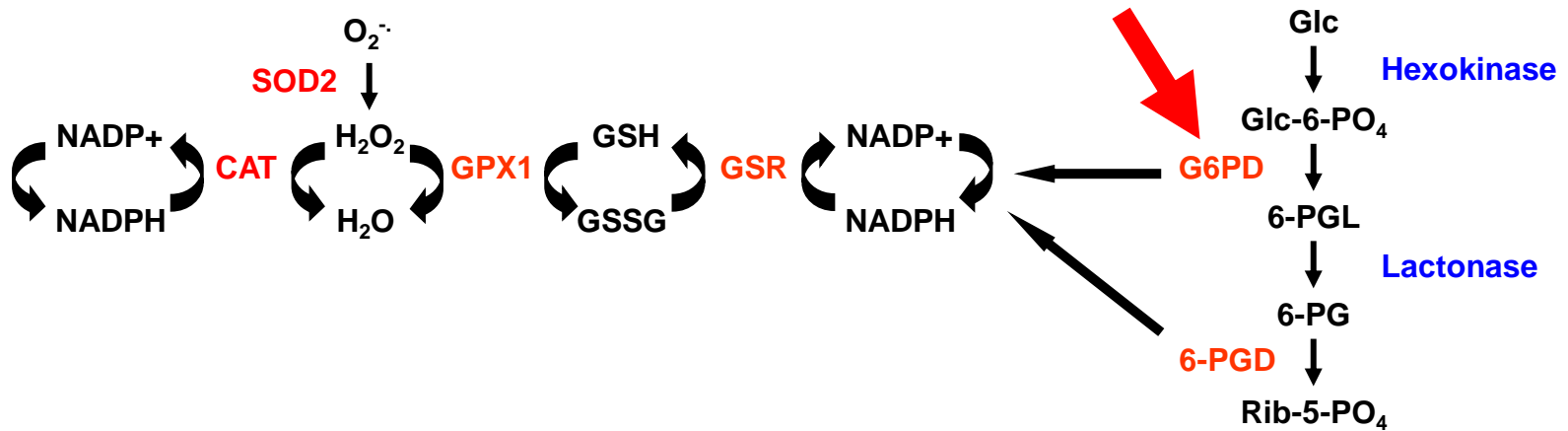


**Decreased RBC recovery *in vivo***

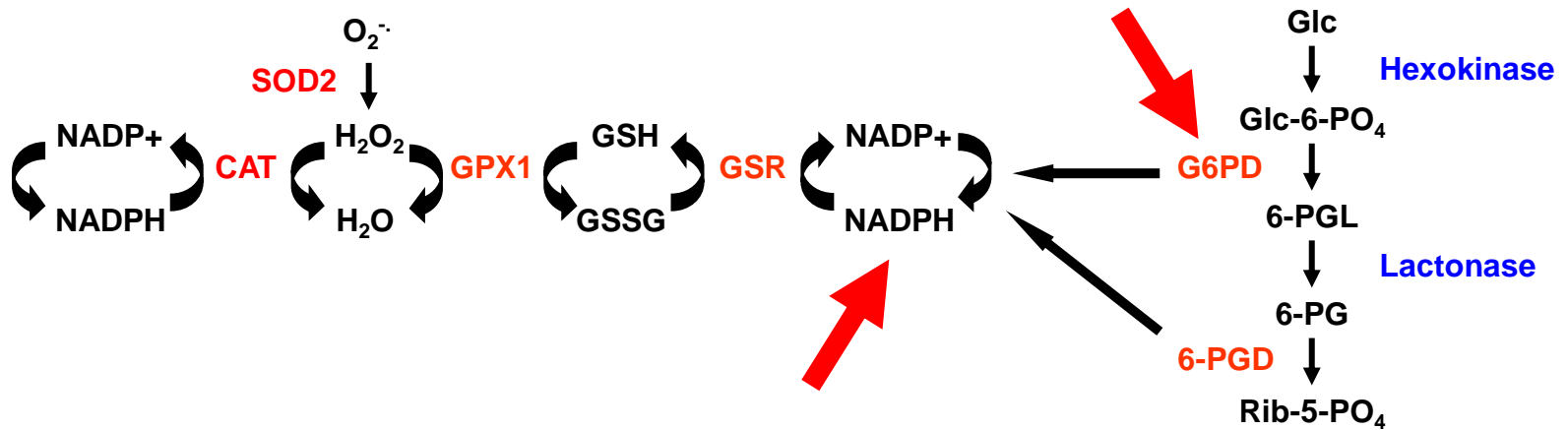
# The case for G6PD: Homeostasis



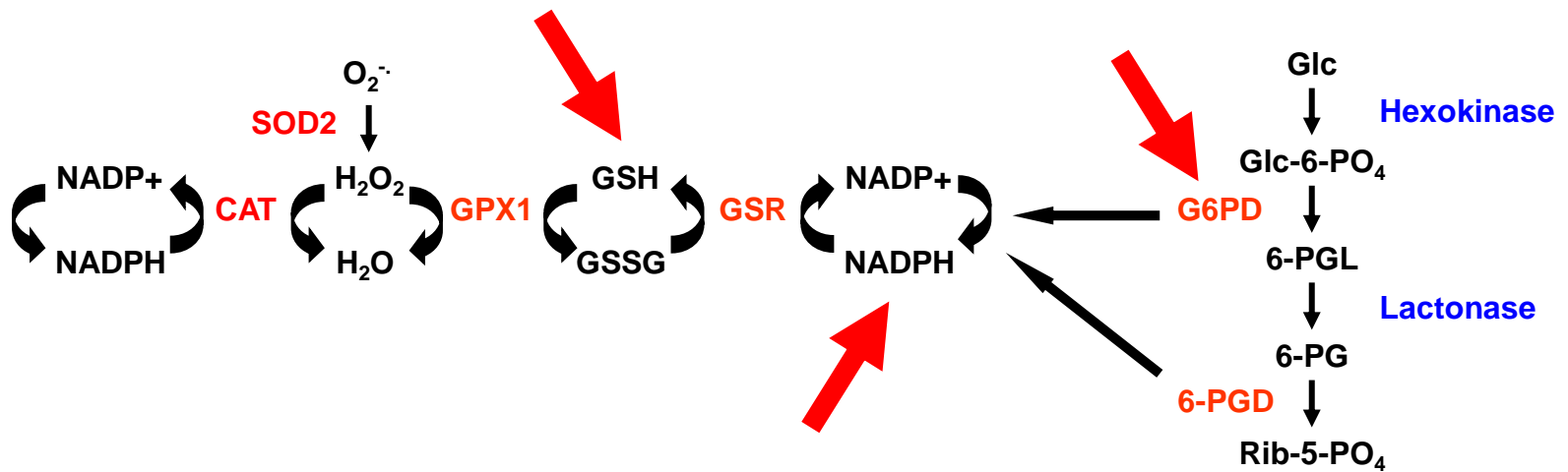
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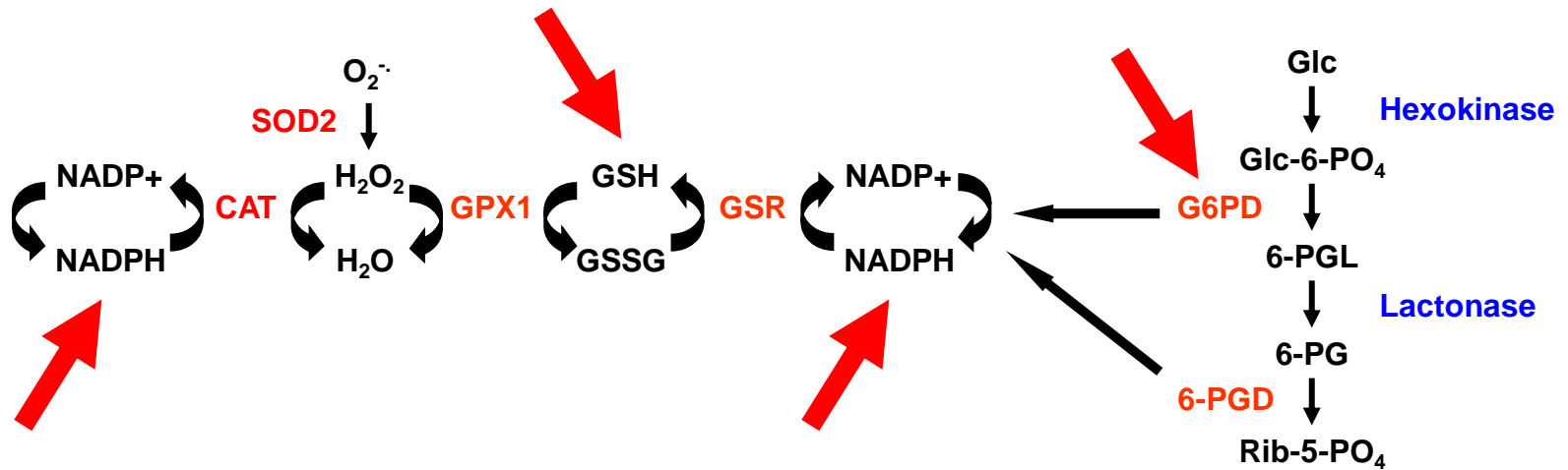


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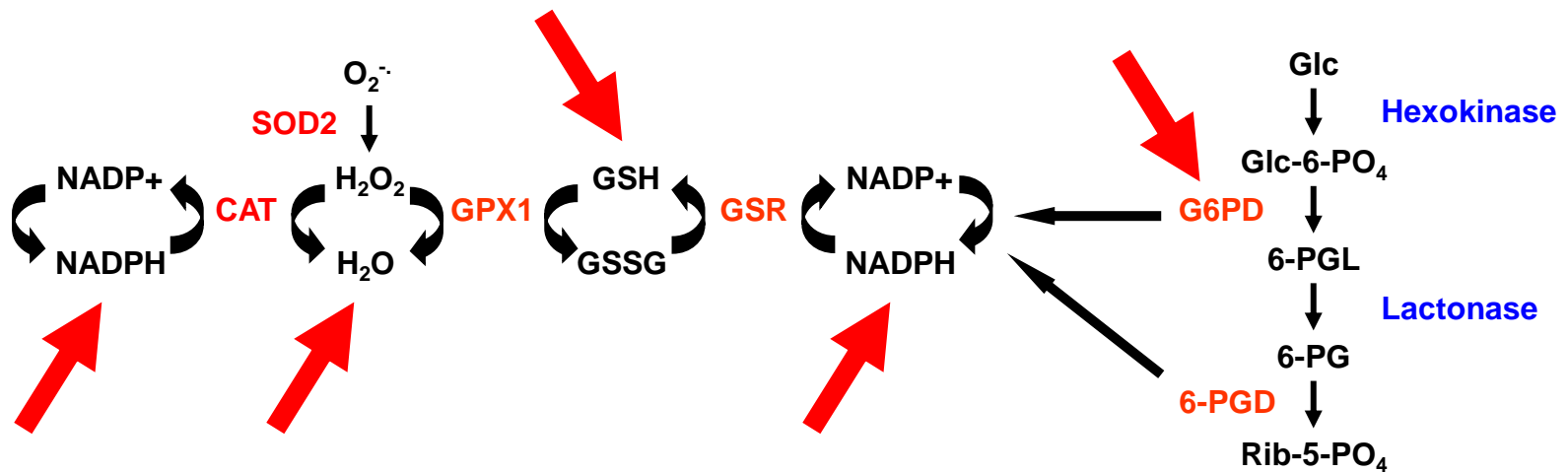




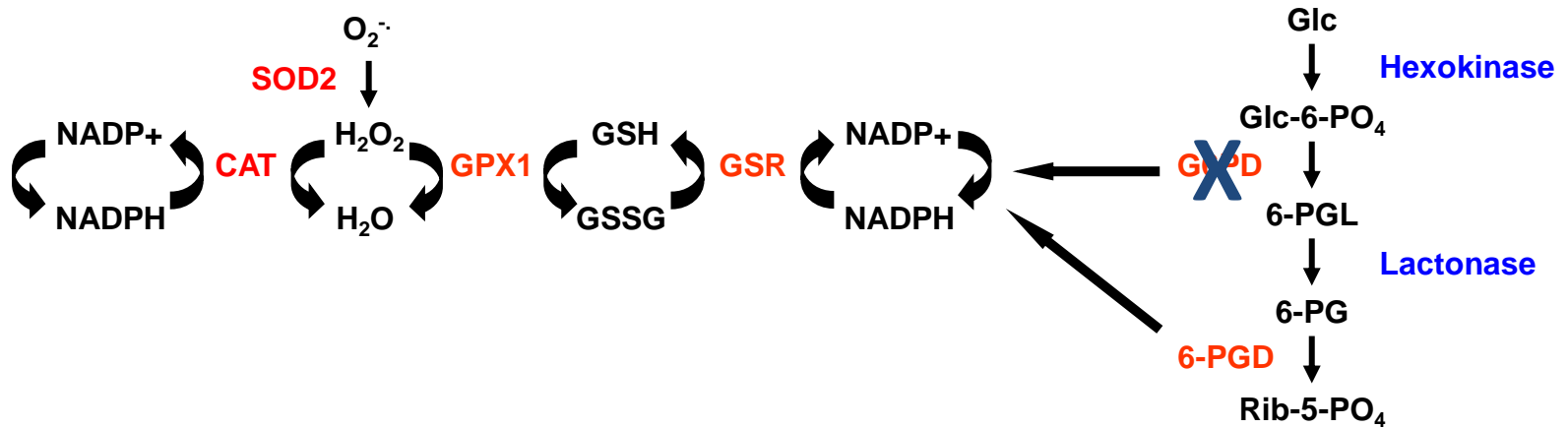
# The case for G6PD: Homeostasis



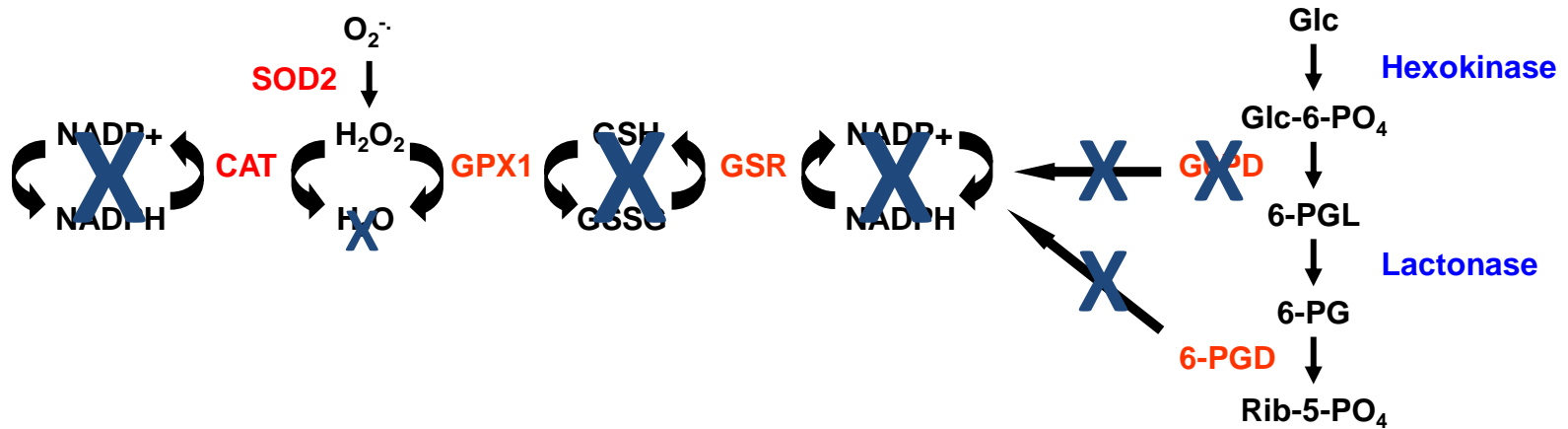
# The case for G6PD: Homeostasis



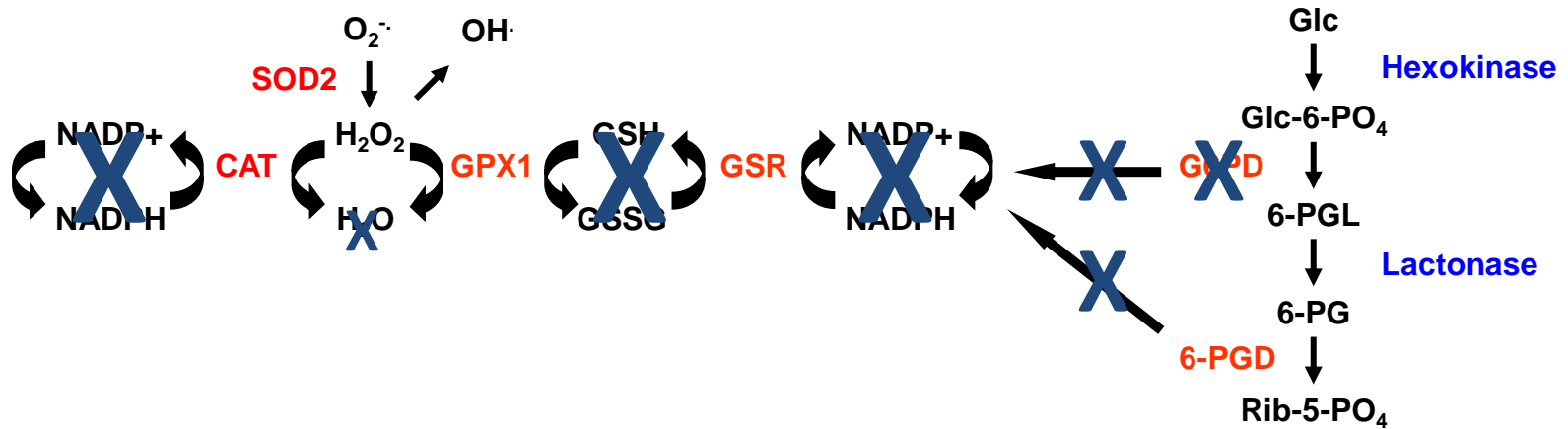
# The case for G6PD: G6PD-deficiency



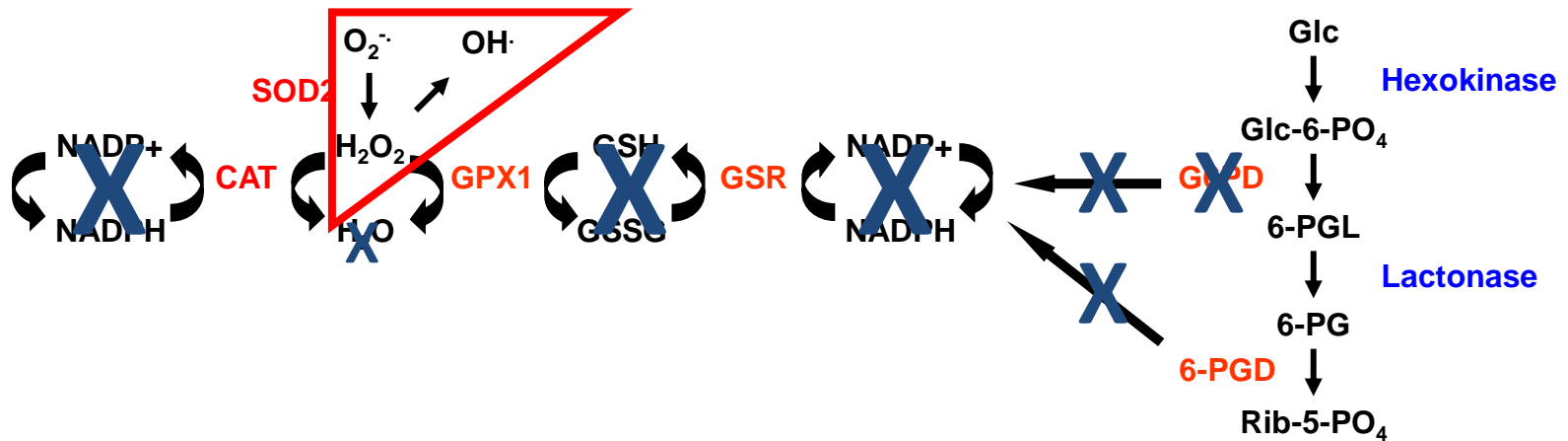
# The case for G6PD: G6PD-deficiency



# The case for G6PD: G6PD-deficiency



# The case for G6PD: G6PD-deficiency



# **The case for G6PD: G6PD-deficiency**

**Unrelieved oxidative stress:**

**RBC structural damage →**

**Intravascular hemolysis (hemoglobinemia)**

**Extravascular hemolysis (NTBI)**

# **The case for G6PD: G6PD-deficiency**

**Most common human enzymopathy**



# **The case for G6PD: G6PD-deficiency**

**Most common human enzymopathy**

**~400 million affected individuals**

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**Severely decreased activity**

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**Increased activity: super storers?**

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**Prevalence of G6PD-deficiency in normal donors  
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**Random donors: 0.3%**

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**Prevalence of G6PD-deficiency in normal donors  
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**Random donors: 0.3%**

**$R_0R_0/R_0r$  donors: 12.3%**

**Exchange Transfusions for Sickle Cell Disease**

# **Study Design**

## **Study plan:**

**10 G6PD-deficient + 30 matched controls**

**Donate 1 unit; pre-storage leukoreduced;  
store for 40-42 days in AS-3; 24h 51-Cr PTR**

# **Study Design**

## **Study plan:**

**10 G6PD-deficient + 30 matched controls**

**Donate 1 unit; pre-storage leukoreduced;**

**store for 40-42 days in AS-3; 24h 51-Cr PTR**

## **Completed study:**

**10 G6PD-deficient + 30 matched controls consented**

**10 G6PD-deficient + 27 controls completed study**

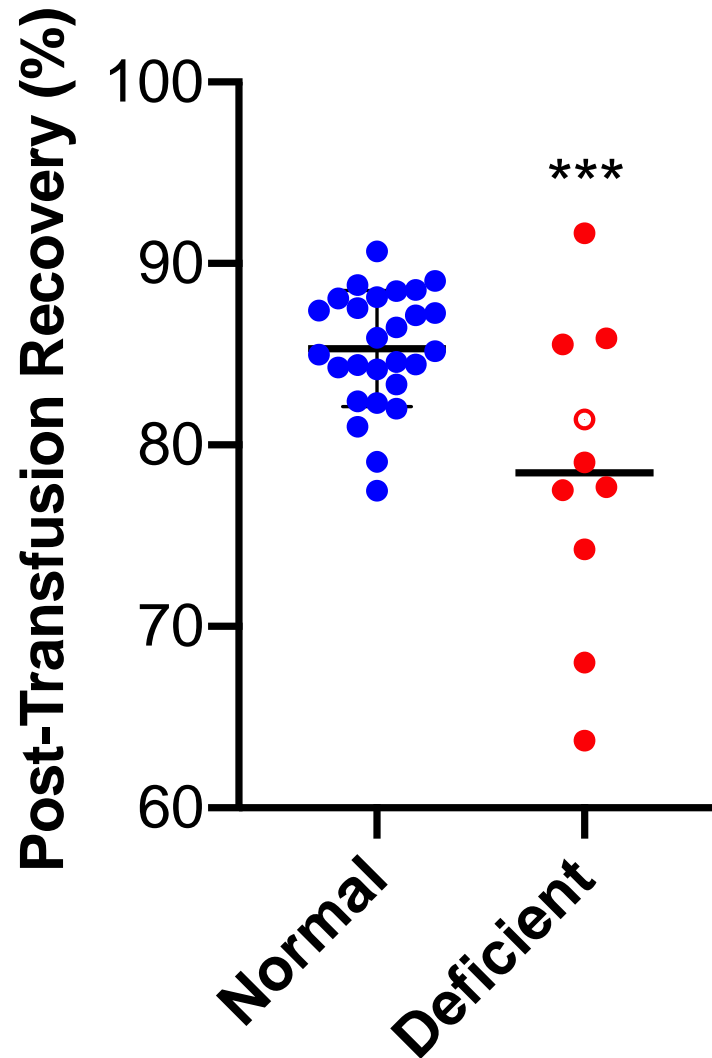
**G6PD-deficient variants: 9 African, 1 Mediterranean**

**Exon sequencing**

**None with hemoglobin variant or thalassemia**

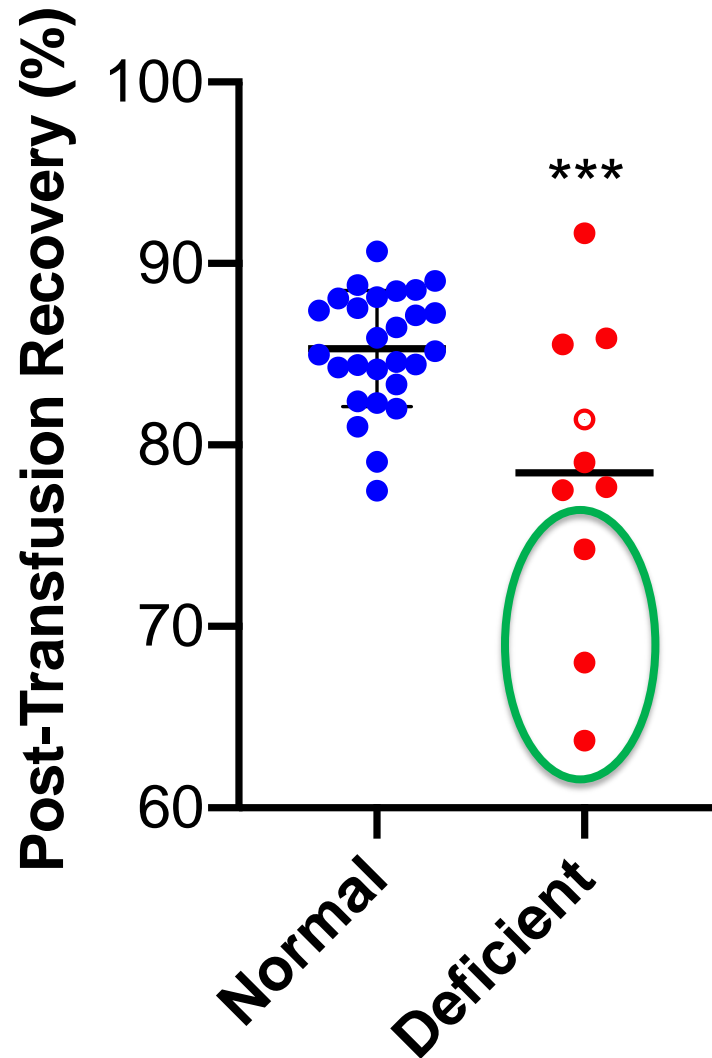
**Hb screen (HPLC) + CBC**

# The case for G6PD: G6PD-deficiency



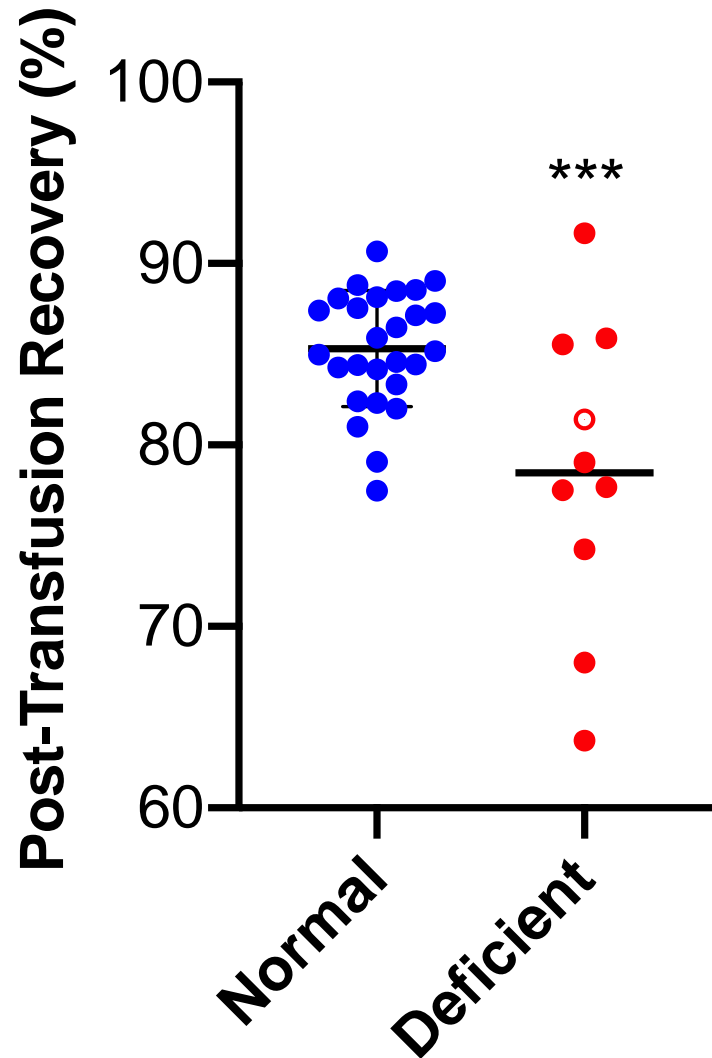
6.8% decreased  
recovery of  
G6PD-deficient  
RBCs

# The case for G6PD: G6PD-deficiency



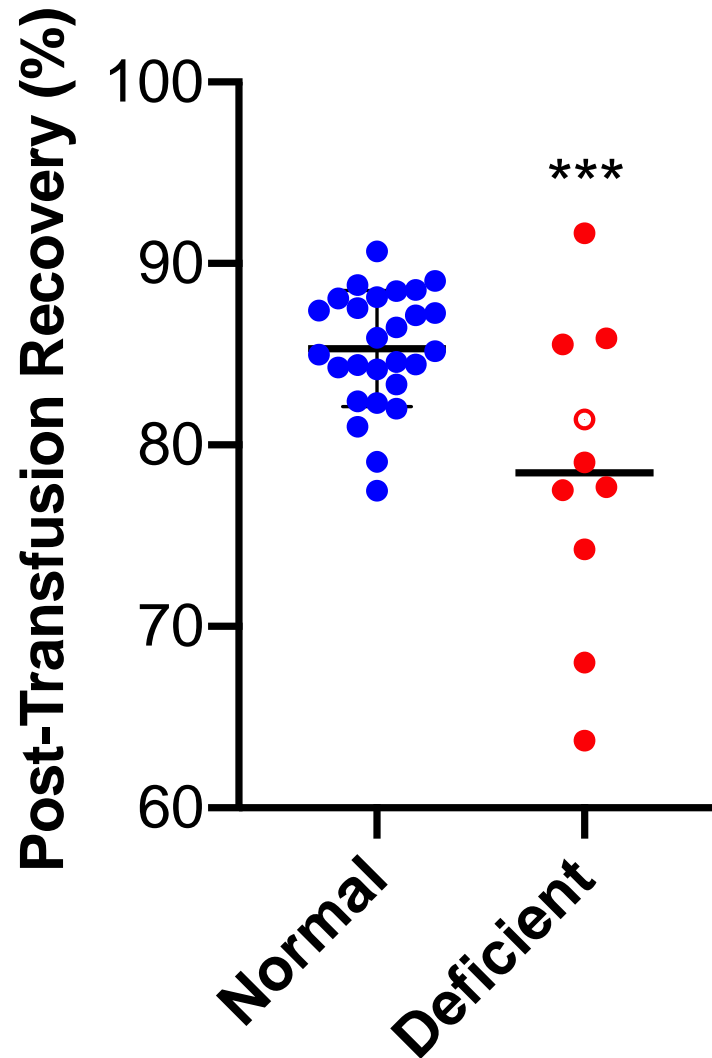
**“FDA failures” in  
G6PD-deficient  
group**

# The case for G6PD: G6PD-deficiency



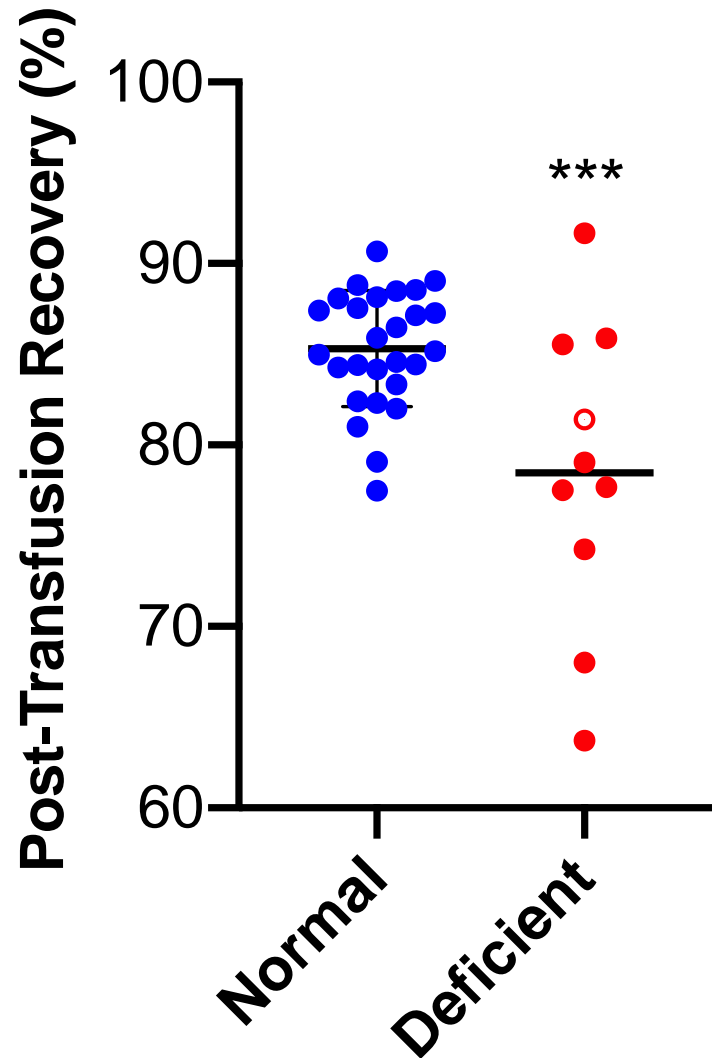
No correlation  
between PTR  
and G6PD  
enzyme activity  
within groups

# The case for G6PD: G6PD-deficiency



No difference  
in hemolysis  
“in the bag” at  
outdate

# The case for G6PD: G6PD-deficiency



Would 24-hour  
post-transfusion  
recoveries be  
worse in ill  
recipients?



# **The case for G6PD:**

## **Next steps**

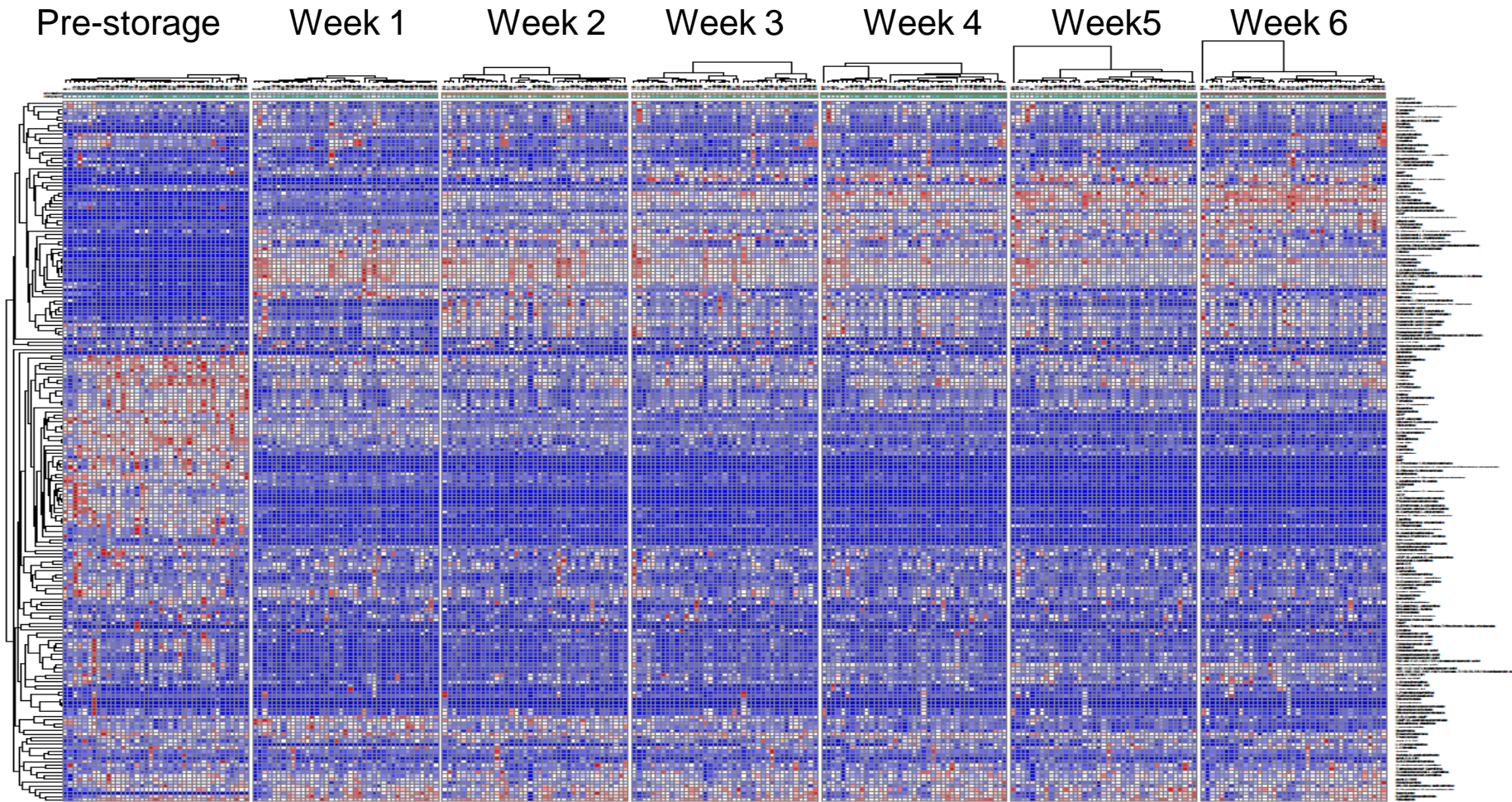
# **The case for G6PD: Next steps**

**Metabolomics:**

**Mechanistic understanding**

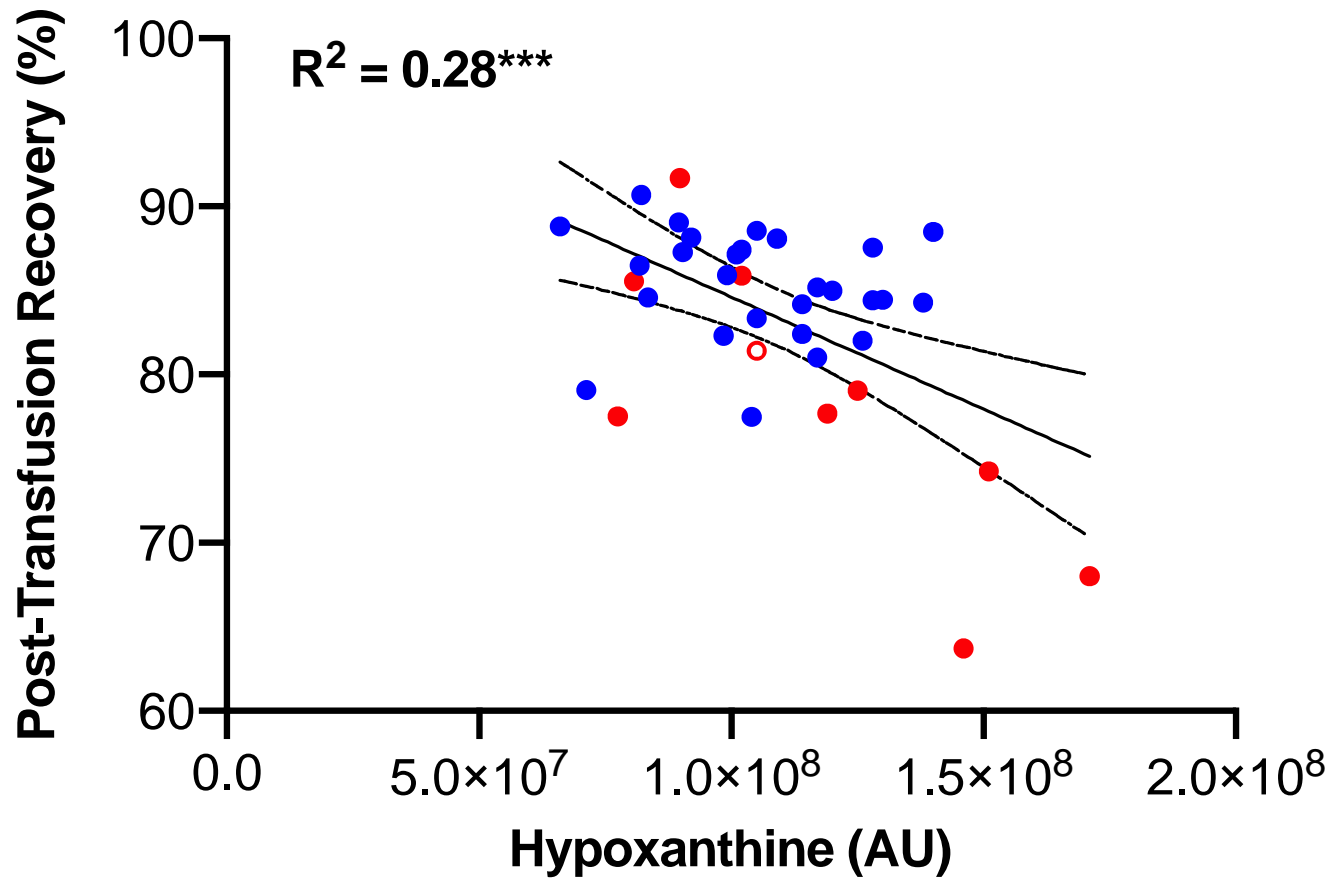
**Develop predictive biosignature(s)**

# The case for G6PD: Next steps



Francis et al. Unpublished observations.

# The case for G6PD: Next steps



Francis et al. Unpublished observations.

# Metabolomics: Human RBCs



**Jim Zimring**



**Angelo D'Alessandro**

# **(Interim) Conclusions**

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**RBCs obtained from G6PD-deficient volunteers have inferior storage quality at 40-42 days**

**Statistically-significant difference of 6.8% ( $p < 0.001$ )**

**Strongly suggests that the RBC's intrinsic ability to resist oxidative stress affects storage quality**

**Is this difference clinically relevant?**

**Acute transfusion setting**

**Chronic transfusion setting**

**Acute intercurrent illness**

**Diet**

**Diet**

**Vitamin C**

**Vitamin E**

**Vitamin D**

**Fe**

**Cu**

**Se**

**Ergothioneine**

**Olive oil & other lipids**

**Obesity**

**Diet**

**Vitamin C**

**Vitamin E**

**Vitamin D**

**Fe**

**Cu**

**Se**

**Ergothioneine**

**Olive oil & other lipids**

**Obesity**

# The “RBC storage lesion”

## Final common pathway?

Metabolic dysfunction & oxidative stress →

↓ Deformability

↑ “Eat me” signals

↓ “Don’t eat me” signals

↑ Hemolysis *in vitro*

↑ RBC clearance *in vivo*

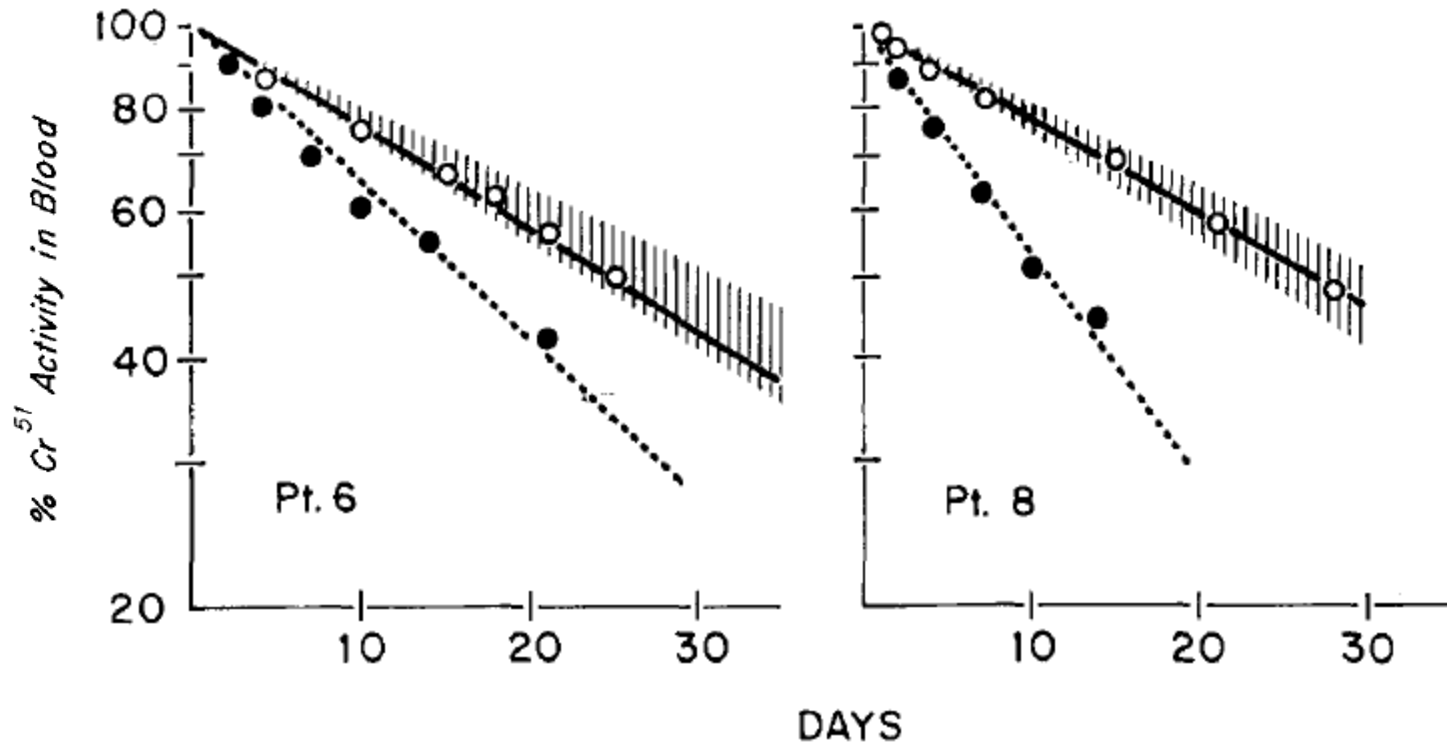
Intravascular and extravascular hemolysis

**Iron deficiency (without anemia) is  
very common in blood donors**

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very common in blood donors**

**Iron-deficient erythropoiesis (IDE)**

# Iron deficiency anemia affects RBC lifespan & transfusion recovery



**Fig. 4.** Cr<sup>51</sup> red cell half-life before (● — ●) and after (○ — ○) correction of iron deficiency anemia.



# Iron deficiency anemia affects RBC lifespan & transfusion recovery

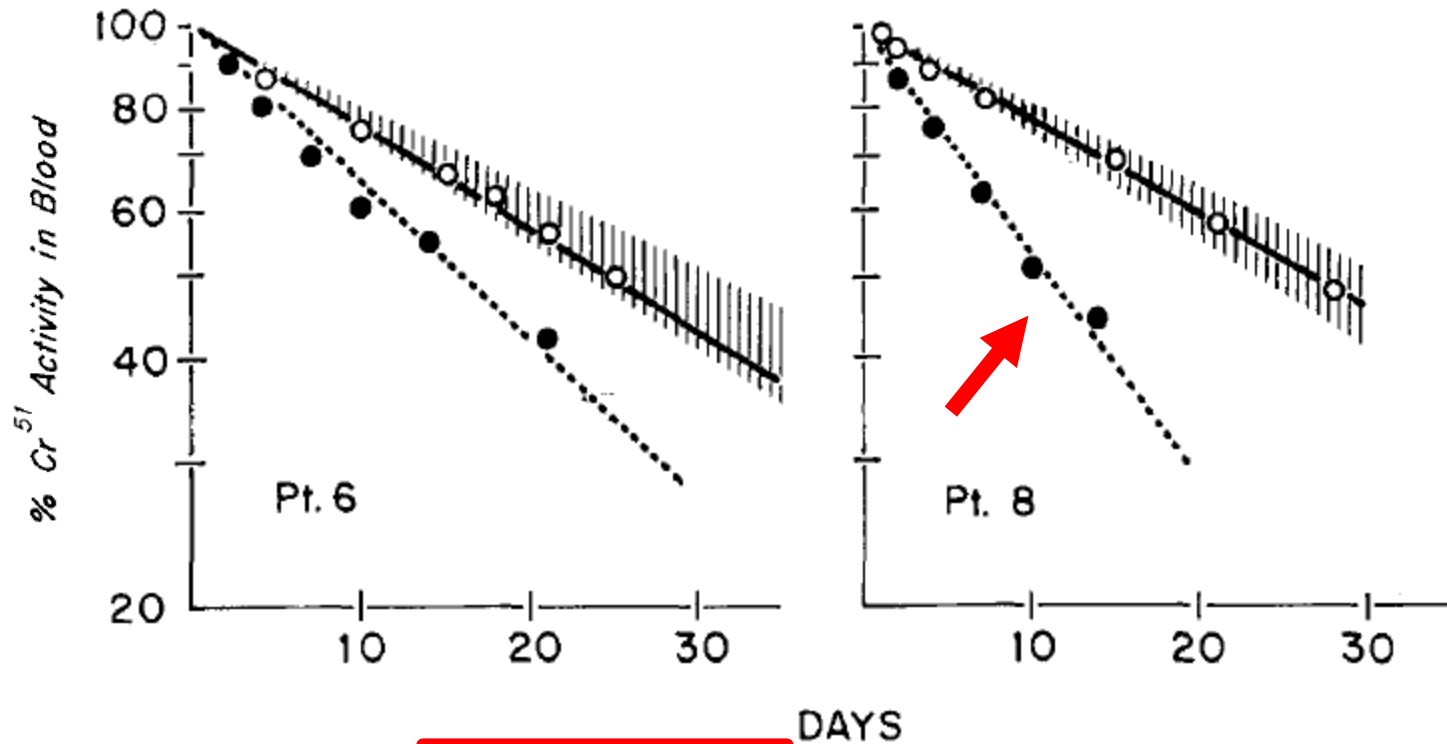
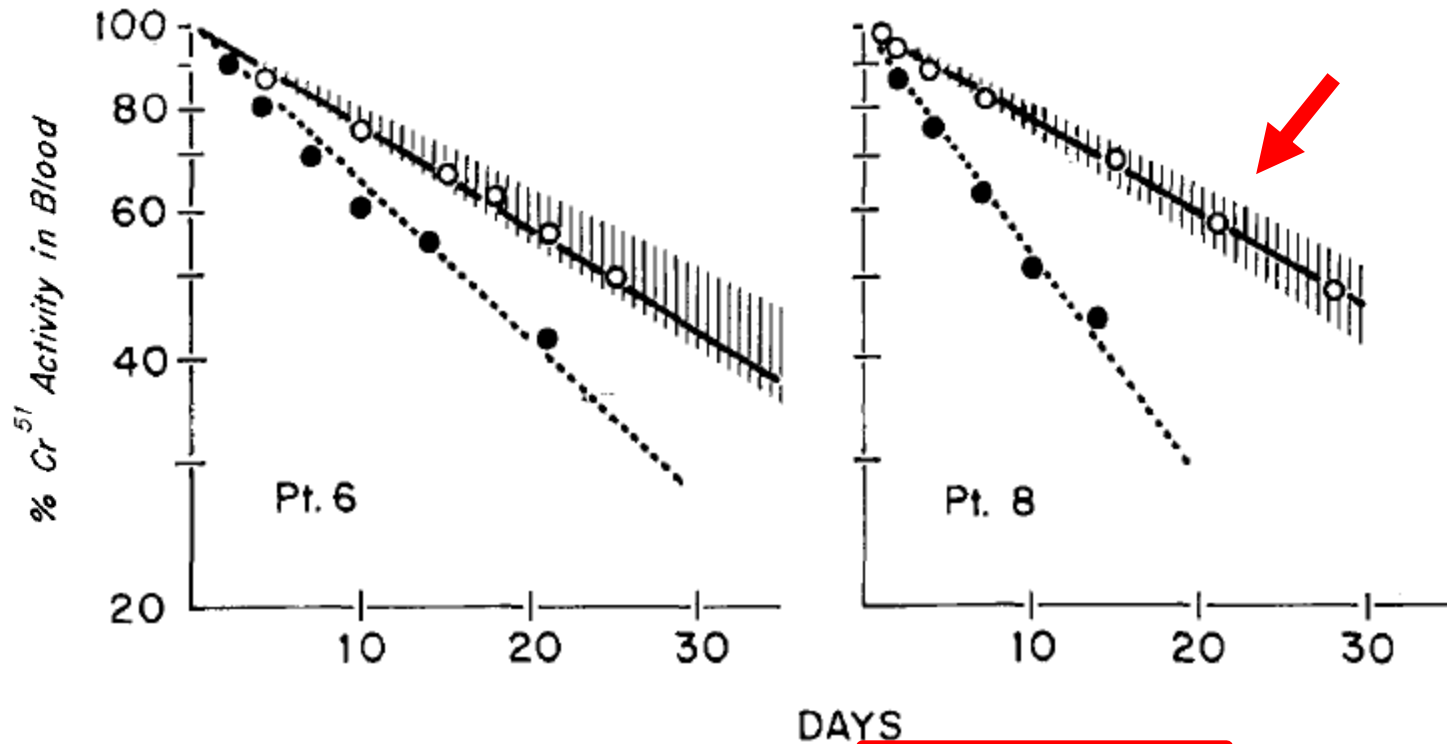


Fig. 4. Cr<sup>51</sup> red cell half-life before (● — ●) and after (○ — ○) correction of iron deficiency anemia.

# Iron deficiency anemia affects RBC lifespan & transfusion recovery



**Fig. 4.** Cr<sup>51</sup> red cell half-life before (● — ●) and after (○ — ○) correction of iron deficiency anemia.

# **Iron deficiency anemia affects RBC lifespan & transfusion recovery**

**↓ Resistance to oxidative stress**

**↑ Oxidative damage**

**↓ Resistance to low pH**

**↑ Phosphatidylserine exposure**

**↓ Deformability**

**↑ Splenic clearance**

# **Iron deficiency anemia affects RBC lifespan & transfusion recovery**

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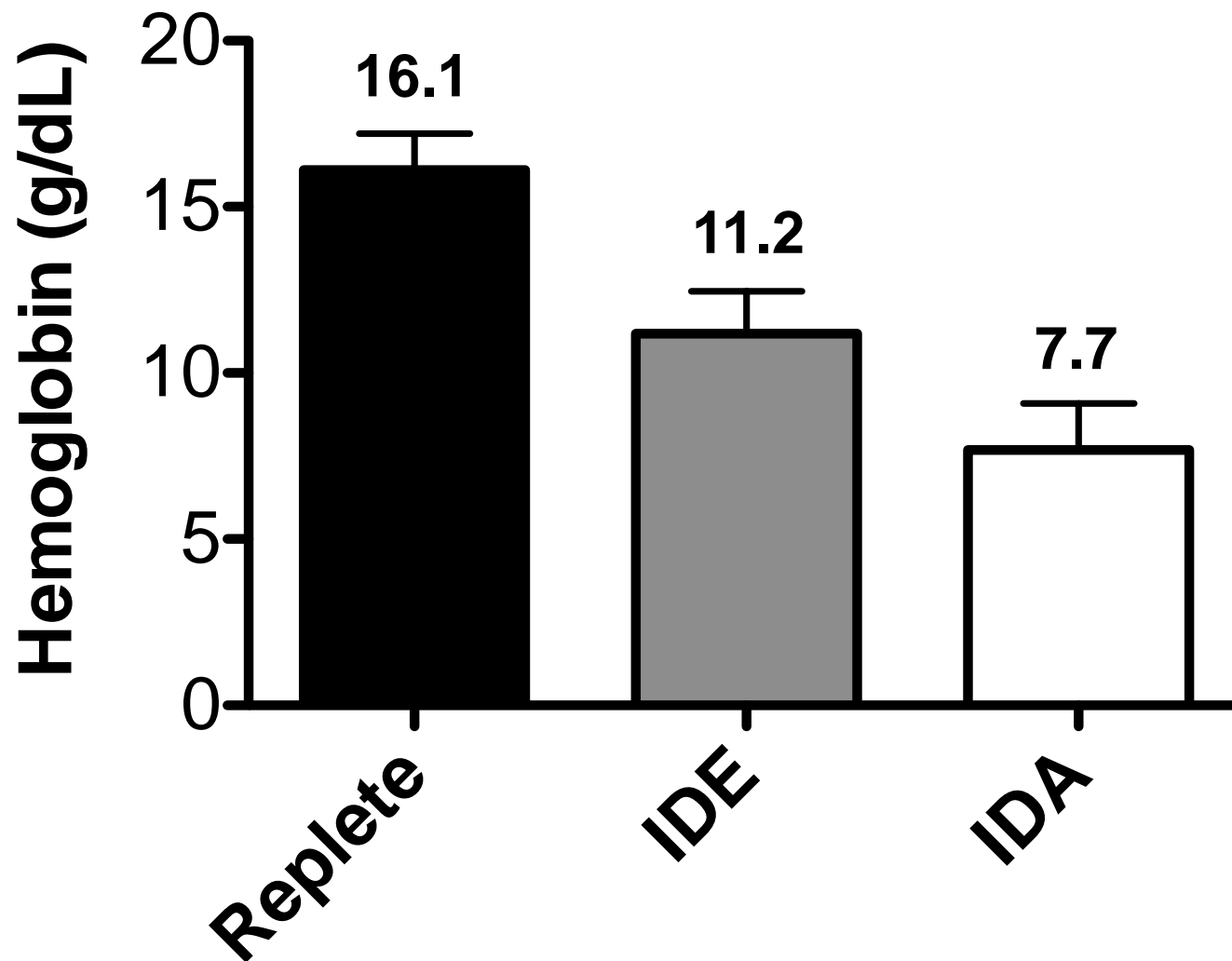
**Do these apply to RBCs in IDE?**

# Mouse model

Weanling, male C57BL/6 mice:

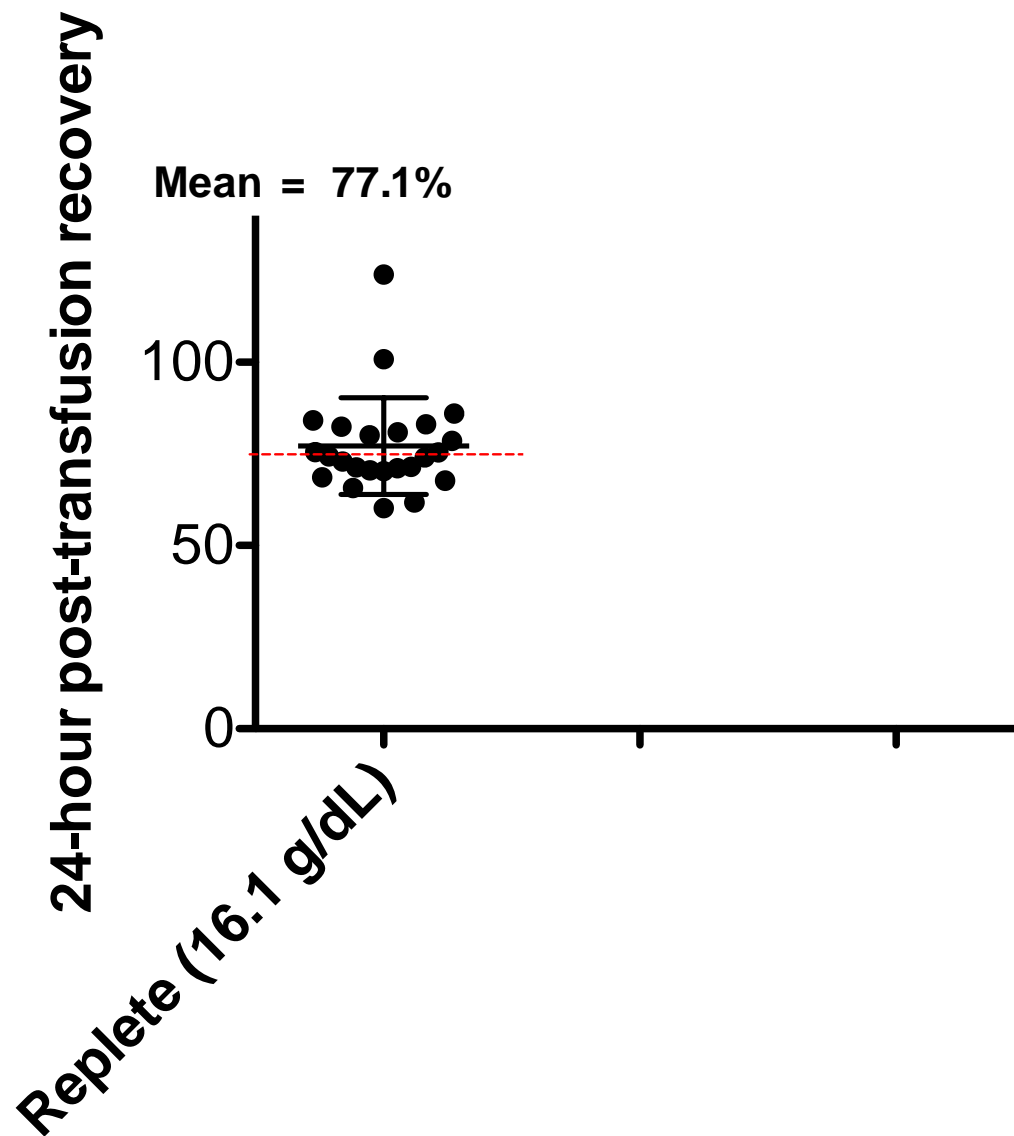
1. Control diet: 45 ppm of iron (normal)
2. Iron-deficient diet: 0-4 ppm of iron (IDE)
3. Iron-deficient diet + weekly phlebotomy (IDA)

# Results

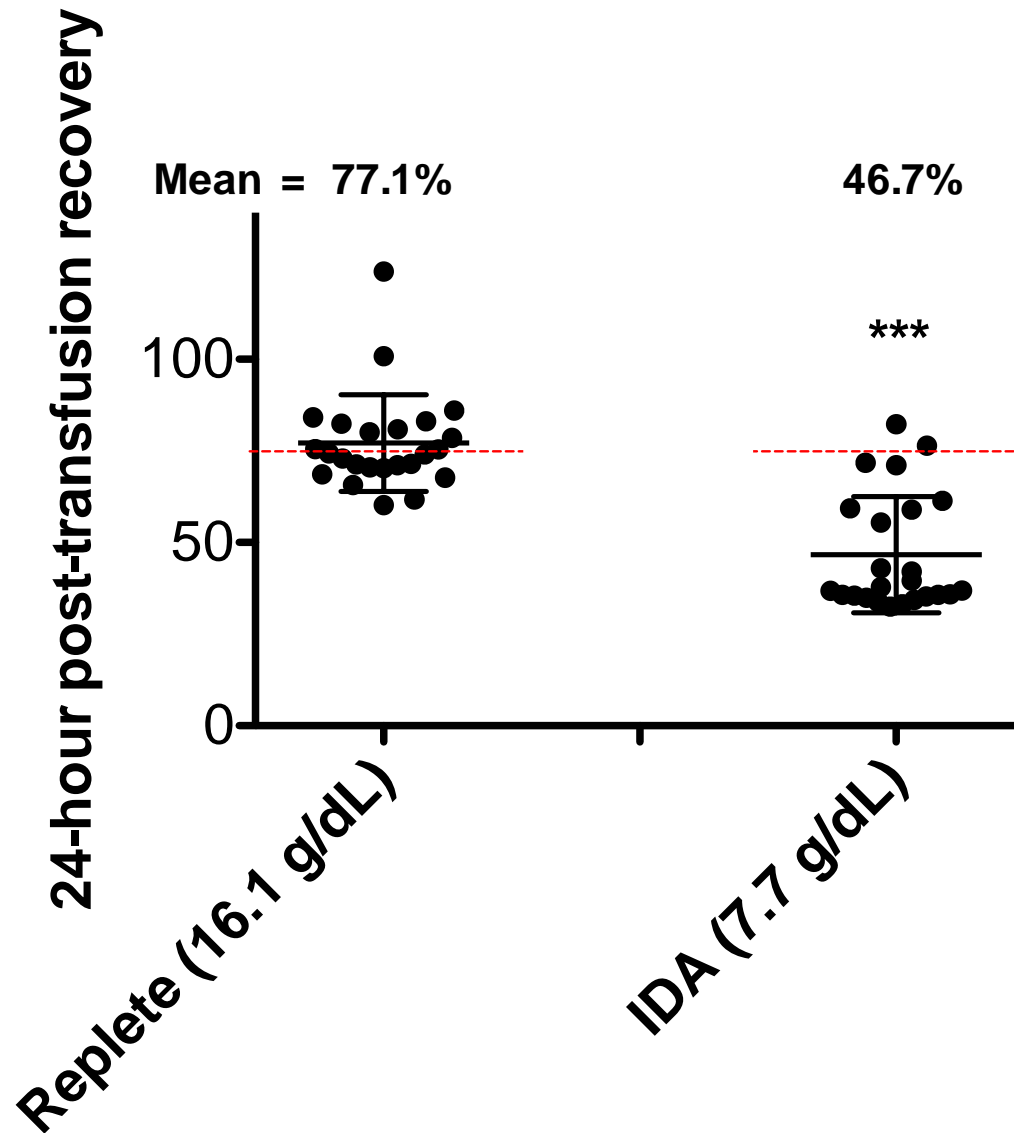


All comparisons:  $p < 0.001$

# Results



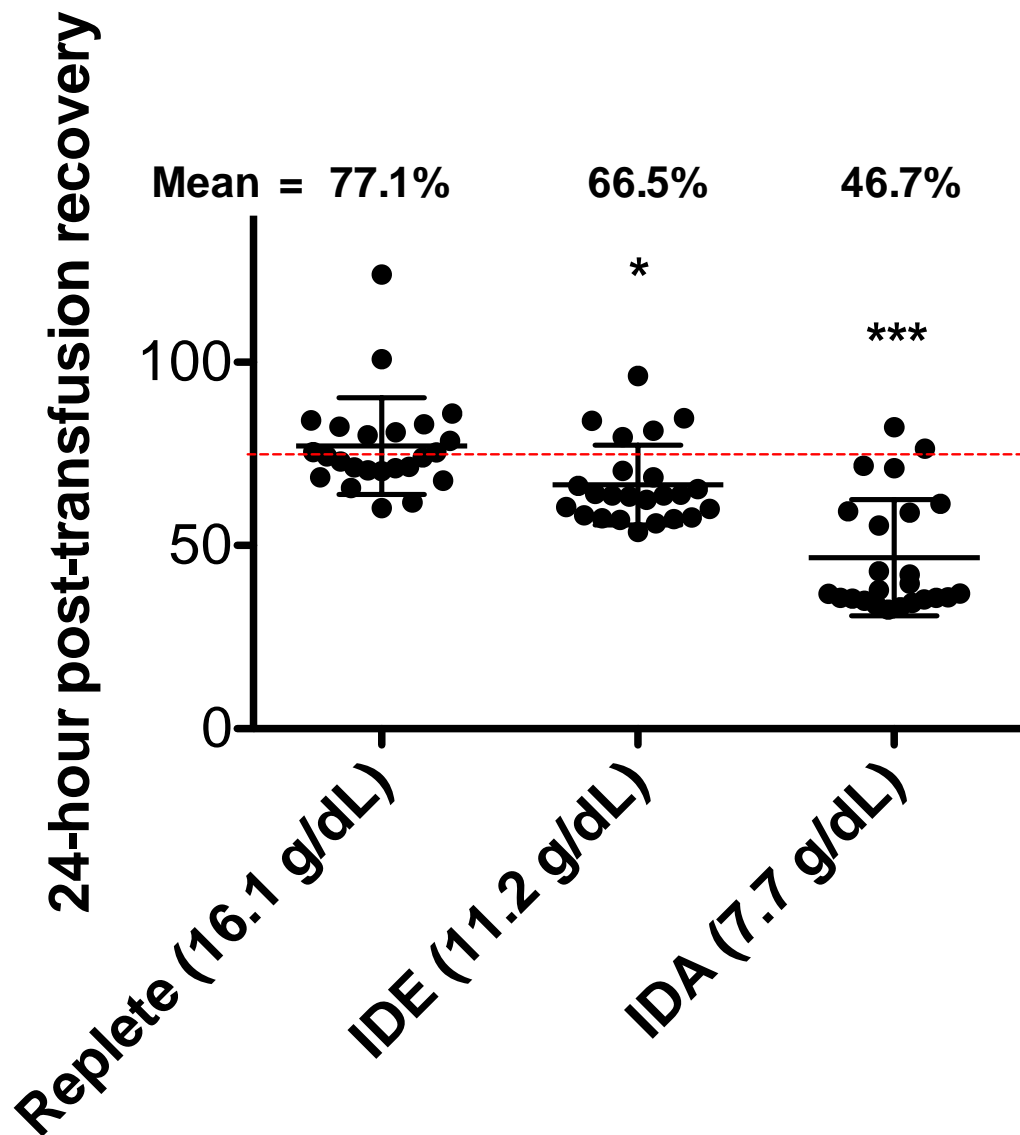
# Results



\*\*\*:  $p < 0.001$



# Results



\*:  $p < 0.05$

# Interim Conclusions

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**RBCs from mice with iron deficiency anemia  
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**RBCs from mice with iron deficiency anemia exhibit poor storage quality**

**RBCs from mice with “iron-deficient erythropoiesis” exhibit suboptimal storage quality**

**Would this be true for human recipients of RBC transfusions from donors with IDE?**

# Would this be true for human recipients of RBC transfusions from donors with IDE?

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## Donor Iron Deficiency Study - Red Blood Cells From Iron-deficient Donors: Recovery and Storage Quality (DIDS)

**This study is enrolling participants by invitation only.**

**Sponsor:**

Columbia University

**Collaborators:**

New York Blood Center  
National Heart, Lung, and Blood Institute (NHLBI)

**Information provided by (Responsible Party):**

Eldad Arie Hod, Columbia University

**ClinicalTrials.gov Identifier:**

NCT02889133

First received: August 25, 2016

Last updated: March 20, 2017

Last verified: March 2017

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# Would this be true for human recipients of RBC transfusions from donors with IDE?

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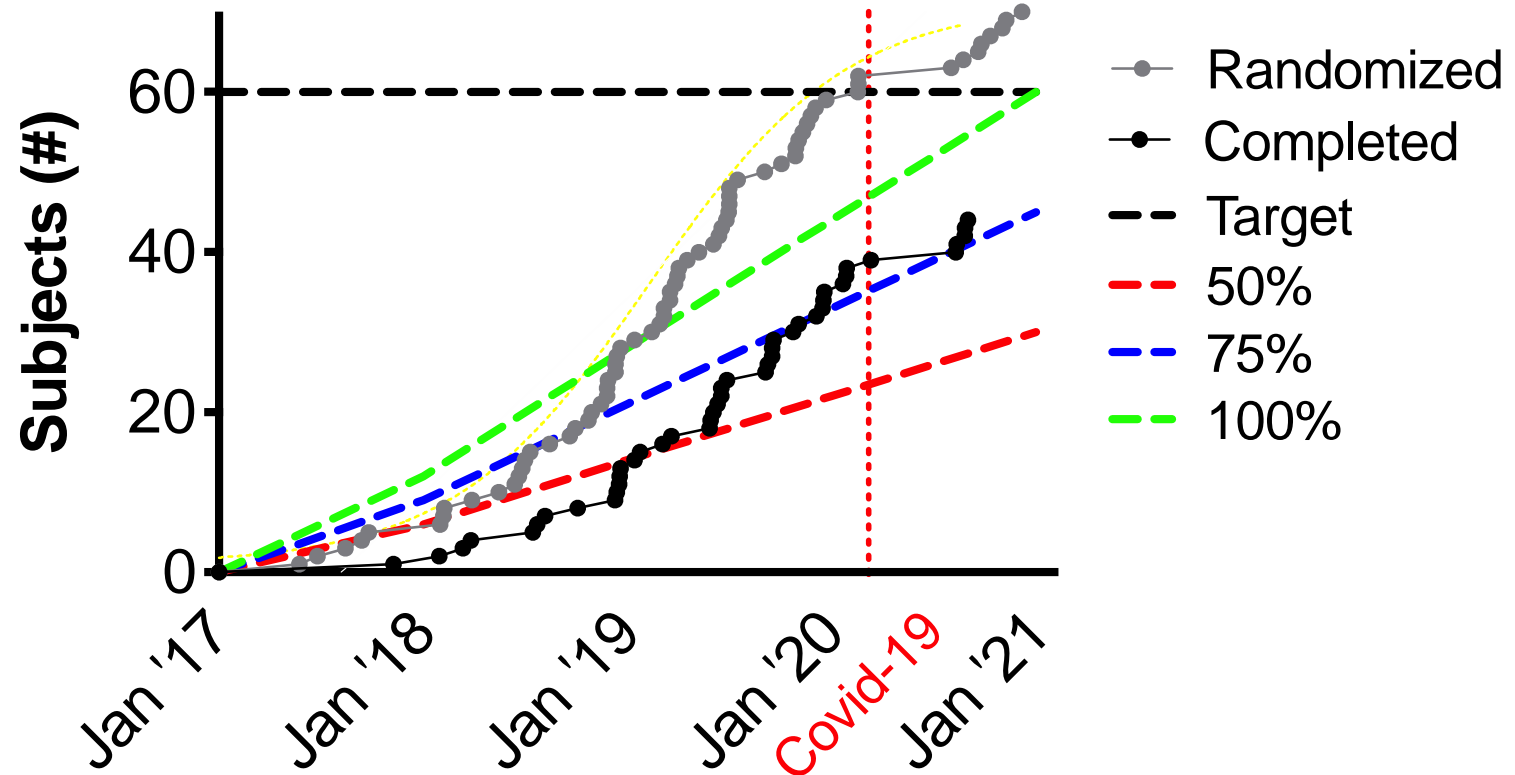
[No Study Results Posted](#)

[Disclaimer](#)

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**E. Hod**

# Would this be true for human recipients of RBC transfusions from donors with IDE?



**All volunteers have been randomized;  
various stages of completion.**



**“Environment”**

# **“Environment”**

**Aging**

**Smoking**

**Microbiome**

**Lead**

**Others?**

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**↑ blood lead level:  $>5 \mu\text{g/dL}$  ( $0.2415 \mu\text{mol/L}$ )**

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**↑ blood lead level:  $>5 \mu\text{g/dL}$  ( $0.2415 \mu\text{mol/L}$ )**

**In whole blood, 75% of lead is in RBCs**

# **Lead**

**Do donor pRBC units contain high lead levels?**

# **Lead**

**Do donor pRBC units contain high lead levels?**

**If so, who cares?**

# Lead

## BLOOD DONORS AND BLOOD COLLECTION

---

### **A population-based study on blood lead levels in blood donors**

*Gilles Delage,<sup>1</sup> Suzanne Gingras,<sup>2</sup> and Marc Rhainds<sup>2,3</sup>*

**TRANSFUSION 2015;55;2633–2640**

**TABLE 2. Characteristics of study participants and estimated GM, 95% CI, and proportion of BLLs of more than 0.15  $\mu\text{mol/L}$  among blood donors population, Québec, Canada 2006 to 2007**

Variable	BLLs					
	Number (%)	GM ( $\mu\text{mol/L}$ )	95% CI	p value	% > 0.15 $\mu\text{mol/L}$	p value
Sex						
Men	2098 (60.1)	0.095	0.038-0.241	<0.001	15.93	<0.001
Women	1392 (39.9)	0.070	0.020-0.242		8.32	
Age (years)						
18-24	655 (9.8)	0.053	0.018-0.154	<0.001	1.28	<0.001
25-34	778 (11.6)	0.060	0.017-0.212		4.64	
35-44	1252 (18.6)	0.075	0.026-0.219		6.07	
45-54	2038 (30.4)	0.095	0.041-0.222		13.62	
55-64	1618 (24.1)	0.122	0.056-0.266		27.44	
$\geq 65$	374 (5.6)	0.135	0.063-0.290		36.59	
Level of education						
Primary school	454 (10.2)	0.089	0.039-0.278	0.008	22.97	<0.001
High school	1466 (32.8)	0.086	0.029-0.253		14.42	
College or university	1566 (57.0)	0.083	0.025-0.231		8.18	
Number of previous blood donations						
0	310 (8.9)	0.092	0.018-0.246	0.025	8.43	<0.001
1-3	598 (17.1)	0.089	0.021-0.225		6.76	
4-10	789 (22.6)	0.092	0.026-0.245		10.88	
11-25	879 (25.2)	0.086	0.033-0.231		11.86	
>25	914 (26.2)	0.088	0.044-0.256		21.55	
Age of dwelling (years)						
<10	354 (10.2)	0.079	0.022-0.236	<0.001	8.32	<0.001
10-29	1139 (32.8)	0.083	0.027-0.228		10.10	
30-49	1161 (33.4)	0.084	0.028-0.248		12.90	
$\geq 50$	818 (23.6)	0.091	0.030-0.274		15.93	
Smoking status						
Never smoker	2540 (73.5)	0.085	0.026-0.240	<0.001	11.30	0.106
Ex-smoker ( $\geq 1$ year)	337 (9.7)	0.091	0.038-0.232		14.78	
Ex-smoker (<1 year)	86 (2.5)	0.107	0.034-0.280		16.44	
Occasional smoker	142 (4.1)	0.091	0.022-0.254		11.22	
Regular smoker (pack-years)	352 (10.2)	0.109	0.033-0.278		14.71	
<5	42	0.093	0.023-0.172	0.219	1.24	<0.001
5-9	36	0.104	0.025-0.248		9.43	
10-19	84	0.099	0.032-0.258		11.92	
20-29	64	0.103	0.044-0.261		12.97	
$\geq 30$	111	0.115	0.058-0.309		30.08	
Alcohol consumption						
Never	355 (10.3)	0.080	0.024-0.257	<0.001	13.19	<0.001
<1/month	536 (15.5)	0.076	0.021-0.229		9.66	
1-3/month	786 (22.7)	0.079	0.024-0.217		8.02	
1-2/week	1100 (31.8)	0.088	0.031-0.237		12.26	
3-6/week	508 (14.7)	0.094	0.038-0.246		13.84	
Daily	171 (5.0)	0.109	0.054-0.309		30.39	
Employed in the past 12 months						
Not	763 (21.9)	0.085	0.034-0.293	<0.001	22.77	<0.001
Not at risk of lead exposure	2584 (75.2)	0.084	0.026-0.223		8.70	
High risk of lead exposure	100 (2.9)	0.129	0.038-0.472		37.11	

# **Lead**

**Highest prevalence:**

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**Frequent blood donors, who are:**



# **Lead**

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# **Lead**

**Highest prevalence:**

**Frequent blood donors, who are:**

**unemployed, uneducated, and older males,**

**who:**

**smoke, drink, and have lived in the same home for a long time.**

# Lead

## **Lead exposure in preterm infants receiving red blood cell transfusions**

Hijab Zubairi<sup>1</sup>, Paul Visintainer<sup>2,3</sup>, Jennie Fleming<sup>1</sup>, Matthew Richardson<sup>1,3</sup> and Rachana Singh<sup>1,3</sup>

Pediatric RESEARCH

Volume 77 | Number 6 | June 2015

# Lead

One transfused aliquot had a lead level of 56 mcg/dl and only one infant received a single transfusion from this aliquot with a total lead load of 7.84 mcg from the single transfusion. This infant's pretransfusion lead level was <1 mcg/dl with the post-transfusion lead level increasing to 9 mcg/dl. The infant's discharge lead level was 1 mcg/dl. This aliquot was not used for any further transfusions and no other infants were exposed to this elevated lead level.

# **Interim Conclusion**

**The effects of donor environmental factors on RBC storage quality have not been well studied**

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## **Blood donor exposome and impact of common drugs on red blood cell metabolism**

**Travis Nemkov,<sup>1,2</sup> Davide Stefanoni,<sup>1</sup> Aarash Bordbar,<sup>3</sup> Aaron Issaian,<sup>1</sup> Bernhard O. Palsson,<sup>4</sup> Larry J. Dumont,<sup>5</sup> Ariel Hay,<sup>6</sup> Anren Song,<sup>7</sup> Yang Xia,<sup>7</sup> Jasmina S. Redzic,<sup>1</sup> Elan Z. Eisenmesser,<sup>1</sup> James C. Zimring,<sup>6</sup> Steve Kleinman,<sup>8</sup> Kirk C. Hansen,<sup>1,2</sup> Michael P. Busch,<sup>9</sup> Angelo D'Alessandro,<sup>1,2</sup> and the Recipient Epidemiology and Donor Evaluation Study III Red Blood Cell–Omics (REDS-III RBC-Omics) Study<sup>10</sup>**

*JCI Insight* 2021;6(3):e146175 <https://doi.org/10.1172/jci.insight.146175>

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**The effects of donor environmental factors on RBC storage quality have not been well studied**

## **Blood donor exposome and impact of common drugs on red blood cell metabolism**

Travis Nemkov,<sup>1,2</sup> Davide Stefanoni,<sup>1</sup> Aarash Bordbar,<sup>3</sup> Aaron Issaian,<sup>1</sup> Bernhard O. Palsson,<sup>4</sup> Larry J. Dumont,<sup>5</sup> Ariel Hay,<sup>6</sup> Anren Song,<sup>7</sup> Yang Xia,<sup>7</sup> Jasmina S. Redzic,<sup>1</sup> Elan Z. Eisenmesser,<sup>1</sup> James C. Zimring,<sup>6</sup> Steve Kleinman,<sup>8</sup> Kirk C. Hansen,<sup>1,2</sup> Michael P. Busch,<sup>9</sup> Angelo D'Alessandro,<sup>1,2</sup> and the Recipient Epidemiology and Donor Evaluation Study III Red Blood Cell–Omics (REDS-III RBC-Omics) Study<sup>10</sup>

# **Future Directions**

**Making better products: Ideal RBC unit**



# **Future Directions**

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**Optimal post-transfusion recovery & lifespan**  
**“Equivalent to fresh”**

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**Optimal post-transfusion recovery & lifespan**  
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Randomized controlled trial of 7, 28, vs 42 day stored red blood cell transfusion on oxygen delivery ( $\text{VO}_2$  max) and exercise duration

Elliott Bennett-Guerrero<sup>1</sup>  | Sabeen Rizwan<sup>1</sup> | Russell Rozensky<sup>1</sup> |  
Jamie L. Romeiser<sup>1</sup> | John Brittelli<sup>1</sup> | Rany Makaryus<sup>1</sup> | Jun Lin<sup>1</sup> |  
Dennis K. Galanakis<sup>1</sup> | Darrell J. Triulzi<sup>2</sup> | Richard E. Moon<sup>3</sup>

**Transfusion 61:699-707, 2021**

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**New screening tests; pathogen reduction/inactivation**

**Thank you**